

1-1-1999

Complementation of conjugation functions of *Streptomyces lividans* plasmid pIJ101 by the related *Streptomyces* plasmid pSB24.2

Gregg S. Pettis
Louisiana State University

Shubha Prakash
Louisiana State University

Follow this and additional works at: https://digitalcommons.lsu.edu/biosci_pubs

Recommended Citation

Pettis, G., & Prakash, S. (1999). Complementation of conjugation functions of *Streptomyces lividans* plasmid pIJ101 by the related *Streptomyces* plasmid pSB24.2. *Journal of Bacteriology*, 181 (15), 4680-4685. <https://doi.org/10.1128/jb.181.15.4680-4685.1999>

This Article is brought to you for free and open access by the Department of Biological Sciences at LSU Digital Commons. It has been accepted for inclusion in Faculty Publications by an authorized administrator of LSU Digital Commons. For more information, please contact ir@lsu.edu.

Complementation of Conjugation Functions of *Streptomyces lividans* Plasmid pIJ101 by the Related *Streptomyces* Plasmid pSB24.2

GREGG S. PETTIS* AND SHUBHA PRAKASH

Department of Biological Sciences, Louisiana State University, Baton Rouge, Louisiana 70803

Received 29 March 1999/Accepted 28 May 1999

A database search revealed extensive sequence similarity between *Streptomyces lividans* plasmid pIJ101 and *Streptomyces* plasmid pSB24.2, which is a deletion derivative of *Streptomyces cyanogenus* plasmid pSB24.1. The high degree of relatedness between the two plasmids allowed the construction of a genetic map of pSB24.2, consisting of putative transfer and replication loci. Two pSB24.2 loci, namely, the *cis*-acting locus for transfer (*clt*) and the transfer-associated *korB* gene, were shown to be capable of complementing the pIJ101 *clt* and *korB* functions, respectively, a result that is consistent with the notion that pIJ101 and the parental plasmid pSB24.1 encode highly similar, if not identical, conjugation systems.

The conjugative plasmid pIJ101 is one of the most extensively studied of the *Streptomyces* extrachromosomal elements. Originally purified from *Streptomyces lividans* ISP5434 (14), pIJ101 (Fig. 1A) is an 8,830-bp (11), circular, high-copy-number plasmid which possesses a broad host range among actinomycete bacteria (14). To produce copies of itself, pIJ101 uses rolling-circle replication (RCR) (6), a process that involves the 456-amino-acid Rep protein encoded by the *rep* gene of pIJ101 (11), a double-strand origin (DSO) of replication where Rep-mediated single-stranded nicking and initiation of first-strand synthesis occurs, and a likely single-strand origin (SSO) termed *sti*, where second-strand synthesis is normally initiated (4). Certain motifs present within its Rep protein, along with conserved sequences present at the site of Rep nicking, have allowed for the classification of pIJ101 as a member of the pIJ101-pJV1 family of RCR plasmids (20), which is comprised mostly of *Streptomyces* replicons (13). The DSO of pIJ101, which has been mapped (3) to a 517-bp region that includes the likely nicking site (20) as well as other sequences required for plasmid replication and copy number control (3), is located between the *rep* gene and *orf56*, a small open reading frame (ORF) of undetermined function (11). The *sti* locus of pIJ101 (4) includes sequences that may form an extensive secondary structure and may contain consensus SSO hexanucleotide base positions (28).

The efficient transfer of pIJ101 is dependent on both the *tra* gene (12, 14), which encodes a *trans*-acting membrane protein with an as yet undetermined function (16), and a *cis*-acting transfer locus (*clt*), which appears to represent a site or sequence that participates in some unknown manner in the transfer event (17). The *clt* locus was originally subcloned on a 142-bp fragment of pIJ101 that spans the 3' end of the *korB* gene, although sequences essential for *clt* function may be present only downstream of the *korB* ORF (17). Although its exact sequence determinants remain to be elucidated, *clt* does not appear to resemble *cis*-acting transfer sequences found on other bacterial plasmids (27), including the only other *Streptomyces* plasmid *clt* function so far identified (19).

The *korB* gene encodes a repressor protein that binds within its own promoter region as well as that of *kilB* (21, 22), a gene

that is involved in, although not absolutely required for, pIJ101 transfer (12). The ability of the KorB protein to repress or override *kilB* is essential, since the unregulated expression of the *kilB* gene on pIJ101 is lethal in *Streptomyces* (12). Like *kilB*, the genes *spdA* and *spdB* contribute to plasmid transfer in an as yet undetermined ancillary manner (11, 12), while similar to *korB* control, the pIJ101 *korA* gene codes for a repressor (21) that appears to negatively regulate the expression of both *korA* and a transcript that includes *tra*, *spdA*, and *spdB* (8).

pSB24.1 is a conjugative plasmid isolated from *Streptomyces cyanogenus* (2), a species notable for its production of the antitumor agent landomycin A (26). Upon introduction of pSB24.1 into *S. lividans*, a stable, transfer-defective, deletion derivative designated pSB24.2 forms (Fig. 1B), which has a reported size of 3,706 bp (2). The deduced Rep protein of pSB24.2 (9) and potential site of Rep-mediated nicking (5) show homology to those of the other members of the pIJ101-pJV1 family of RCR plasmids (13). Here we show that a database search for sequences showing homology to the pIJ101 *clt* region (and thus plasmids that may share the same transfer mechanism as pIJ101) subsequently revealed extensive similarity between both transfer and replication loci of pIJ101 and corresponding previously unidentified loci on pSB24.2. We further show that the *clt* and *korB* functions encoded by pSB24.2 can complement their pIJ101 counterparts. These results support the hypothesis that pIJ101 and pSB24.1 share the same conjugation system and that pIJ101-like conjugative plasmids may be dispersed throughout at least a portion of the *Streptomyces* genus.

pIJ101 and pSB24.2 show extensive sequence similarity. By using the BLASTN algorithm (1) and the 142-bp *clt*⁺ region of plasmid pIJ101 (Fig. 2) as a query sequence, a database search revealed only one other highly similar sequence, which belonged to *Streptomyces* plasmid pSB24.2. Subsequent BLASTN alignment of the entire pIJ101 and pSB24.2 sequences revealed extensive similarity between them (i.e., approximately 77% of the pSB24.2 sequence aligned to pIJ101 with 89% identity and 1% gaps by this method) and allowed the identification of putative transfer and replication loci on pSB24.2 (Fig. 1B) as detailed below. To resolve discrepancies arising through probable errors, both strands of the *korB-clt* region of pSB24.2 were sequenced by using relevant primers and an ABI Prism dye terminator cycle sequencing ready reaction kit (PE Applied Biosystems, Inc., Foster City, Calif.), followed by anal-

* Corresponding author. Mailing address: Department of Biological Sciences, Louisiana State University, 508 Life Sciences Building, Baton Rouge, LA 70803. Phone: (225) 388-2798. Fax: (225) 388-2597. E-mail: gpettis@unix1.sncc.lsu.edu.

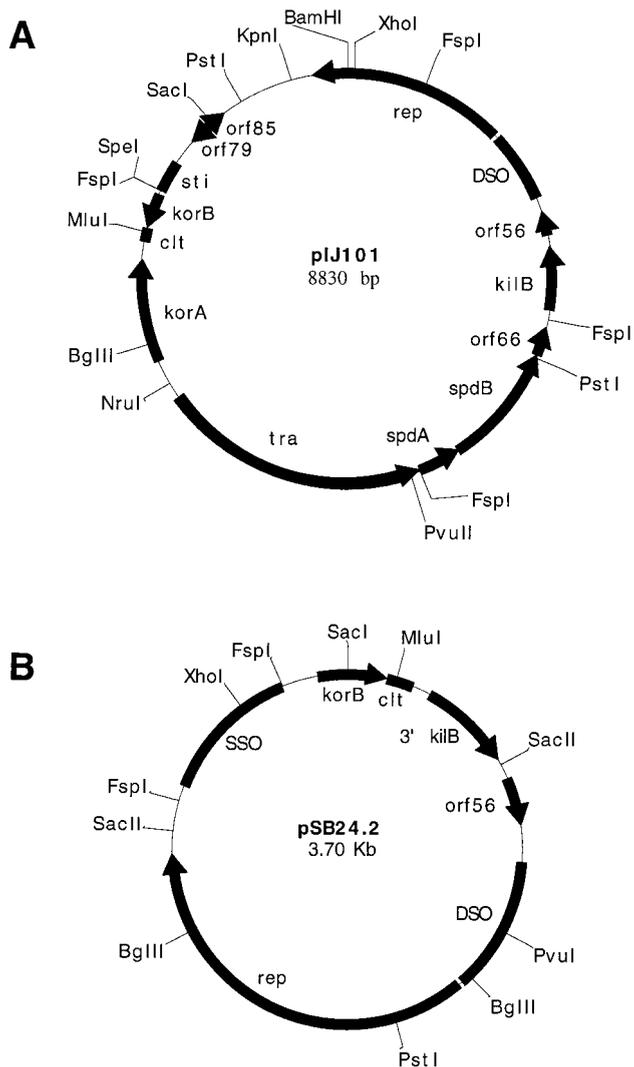


FIG. 1. Physical and genetic maps of pIJ101 (A) and pSB24.2 (B). Filled arrows represent ORFs which were determined from the published pIJ101 (11) or pSB24.2 (2) sequence or from the pSB24.2 sequence reported here. Genetic functions were determined for either pIJ101 genes (12, 14) or loci (solid bars with no arrows) (4, 17) as described previously. Small pIJ101 ORFs of undetermined genetic function (i.e., *orf56*, *orf66*, *orf79*, and *orf85*) are indicated. Certain pSB24.2 genetic functions (i.e., *korB* and *clt*) were determined here; other genetic functions (i.e., DSO and SSO) or small ORFs of unknown function (i.e., *orf56*) were assigned based on their similarity to corresponding pIJ101 sequences or in the case of the *rep* gene on the similarity of the deduced Rep protein to Rep proteins of other *Streptomyces* plasmids (9, 13).

ysis of reactions on a model 310 genetic analyzer (PE Applied Biosystems, Inc.).

The alignment of the revised *korB-clt* sequence of pSB24.2 with the corresponding region of pIJ101 revealed several features (Fig. 2). The 240-bp *korB* ORFs of the two plasmids are 97.5% identical, and the putative 80-amino-acid pSB24.2 KorB protein is identical to the pIJ101 KorB protein (11), with the exception of a histidine residue in place of a glutamine at amino acid position 3 and a lysine residue instead of a glutamic acid at position 30. The latter amino acid position is within the possible DNA binding domain of the mature pIJ101 KorB repressor protein (11, 24), although not at one of the most conserved positions of this motif (15). A comparison of the



FIG. 2. Alignment of the *korB-clt* regions of plasmids pIJ101 and pSB24.2. The revised *korB-clt* sequence of pSB24.2 is shown, aligned with the previously described *korB-clt* region of pIJ101 (11). One-letter amino acid designations are shown above (for pIJ101) or below (for pSB24.2) the corresponding *korB* ORF. Amino acid differences between the pIJ101 KorB protein and the deduced KorB protein of pSB24.2 are in bold. Putative -35 and -10 promoter determinants for the pIJ101 *korB* gene, which are based on previous transcriptional mapping (22), are indicated. Inverted repeats on both sequences are designated by arrows. The 142-bp region of pIJ101 that is known to contain *clt* activity (17) and that was used as a query sequence here for database searches is indicated.

promoter region of the pIJ101 *korB* gene (which was identified following the demonstration that transcription initiates at the A residue of the start codon) (22) with the region immediately upstream of the pSB24.2 *korB* gene revealed that pSB24.2 contains an identical potential -10 but not a completely conserved possible -35 promoter determinant. Also within this region of pSB24.2 are inverted repeat sequences that are identical to those found in the pIJ101 *korB* promoter which were shown previously to be important for the autoregulation of *korB* gene expression (25). Downstream of the *korB* ORFs on both plasmids, there is complete conservation of sequences in the immediate vicinity of the *MluI* restriction site, a location at

TABLE 1. Bacterial strains and plasmids used in this study

Strain or plasmid	Description ^a	Source or reference
<i>S. lividans</i>		
TK23	<i>spc-1</i>	7
TK23.42	TK23 <i>tra</i> and <i>korA</i> genes of pIJ101 chromosomally integrated	17
<i>Escherichia coli</i> DH10B	F ⁻ <i>mcrA</i> Δ(<i>mrr-hsdRMS-mcrBC</i>) φ80 <i>dlacZ</i> ΔM15 Δ <i>lacX74 deoR recA1 endA1 araD139</i> Δ(<i>ara leu</i>)7697 <i>galU galK</i> λ ⁻ <i>rpsL nupG</i>	Gibco BRL
Plasmids		
pGSP149	pIJ350 derivative with pSP72 at the <i>PstI</i> site	17
pGSP263	pIJ101 Clt ⁺ derivative of pGSP149	17
pGSP290	KilB ⁺ derivative of pSP72 containing the 1.0-kb <i>PstI-BalI</i> region of pIJ101 cloned as a blunt-end fragment in the <i>EcoRV</i> site (<i>kilB</i> is transcribed towards the T7 promoter)	This study
pGSP312	pSP72 derivative containing the 142-bp Clt ⁺ region of pSB24.2 cloned as a <i>BamHI</i> fragment in the <i>BamHI</i> site	This study
pGSP313	pIJ350 derivative containing pGSP312 at the <i>PstI</i> site	This study
pGSP317	Derivative of pHYG1 with pGSP290 at the <i>PstI</i> site (pIJ101 <i>rep</i> gene is transcribed towards the SP6 promoter)	This study
pGSP321	KorB ⁺ derivative of pGSP317 containing the 0.3-kb <i>SpeI-MluI</i> region of pIJ101 cloned as a blunt-end fragment in the <i>PvuII</i> site (<i>korB</i> is transcribed towards the SP6 promoter)	This study
pGSP322	KorB ⁺ derivative of pGSP317 containing the 0.4-kb <i>FspI-MluI</i> region of pSB24.2 cloned as a blunt-end fragment in the <i>PvuII</i> site (<i>korB</i> is transcribed towards the SP6 promoter)	This study
pHYG1	Conjugally deficient Hyg ^r derivative of pIJ101 lacking all transfer functions	12
pIJ101	Conjugative <i>S. lividans</i> plasmid	14
pIJ350	Conjugally deficient Tsr ^r derivative of pIJ101 lacking all transfer functions	14
pSB24.2	Conjugally deficient Clt ⁺ KorB ⁺ deletion derivative of <i>S. cyanogenus</i> plasmid pSB24.1	2; obtained from the Russian National Collection of Industrial Microorganisms
pSB24.202	Tsr ^r derivative of pSB24.2 with the 1.0-kb <i>tsr</i> gene (7) cloned as an <i>XhoI</i> fragment at the <i>XhoI</i> site (<i>tsr</i> gene transcribed away from <i>korB</i>)	This study
pSP72	Ap ^r <i>E. coli</i> cloning vector	Promega Biotech

^a Hyg^r, Tsr^r, and Ap^r indicate hygromycin, thiostrepton, and ampicillin resistance, respectively.

unknown, undergoes a spontaneous deletion of sequences (including its transfer-essential *tra* gene) that correspond to the portion of pIJ101 extending from within *kilB* clockwise to just beyond the *korA* gene (Fig. 1A); such a deletion would thereby result in the formation of the transfer-defective pSB24.2 derivative (Fig. 1B).

pSB24.2 encodes a functional *clt* locus that complements the pIJ101 *clt* locus. Our interest in conjugation in *Streptomyces* prompted us to examine whether pSB24.2 encodes a functional *clt* locus, and if so, whether this pSB24.2 locus can complement the *clt* function of the closely related pIJ101 plasmid. Such complementation could potentially allow for the subsequent identification of conserved functional sequences that are important for *clt* activity. To test these possibilities, we used a previously described mating system (16, 17) involving an *S. lividans* donor strain (TK23.42), which contains chromosomally integrated copies of the pIJ101 *tra* and *korA* genes, and an

S. lividans recipient strain (TK23) containing pHYG1 (12), a transfer-defective, hygromycin-resistant pIJ101 derivative. TK23.42 was transformed with pSB24.202 (Table 1), a thiostrepton-resistant derivative of pSB24.2, and plasmid-containing spores of this strain were mixed with TK23(pHYG1) recipient spores, plated on nonselective MY (malt extract-yeast extract) agar (17), and then quantified following mating for donor, recipient, and transconjugant cell types as described previously (17).

As shown in Table 2, pSB24.202 transferred at a frequency (28%) that was nearly identical to that seen for a Clt⁺ pIJ101 derivative (i.e., pGSP263 at 29%) and that was 35-fold higher than the transfer frequency (0.80%) observed for pGSP149, a pIJ101-derived plasmid that lacks *clt*. This result strongly suggested that a *clt* function present on pSB24.2 can function in conjunction with the pIJ101 *tra* gene to promote the efficient transfer of pSB24.2. To show directly that the suspected

TABLE 2. Transfer frequencies of pSB24.2 Clt⁺ derivatives from *S. lividans* TK23.42^a

Plasmid	Donor (relevant genotype)	Recipient	Avg no. of:			Transfer frequency ^b (%)
			Donors	Recipients	Transconjugants	
pGSP263 (<i>clt</i> ⁺ [pIJ101])		pHYG1	3.8 × 10 ⁵	2.1 × 10 ⁷	1.1 × 10 ⁵	29
pGSP149 (<i>clt</i>)		pHYG1	9.0 × 10 ⁴	1.1 × 10 ⁷	7.2 × 10 ²	0.80
pSB24.202 (<i>clt</i> ⁺ [pSB24.2])		pHYG1	9.7 × 10 ⁵	3.9 × 10 ⁷	2.7 × 10 ⁵	28
pGSP313 (<i>clt</i> ⁺ [pSB24.2])		pHYG1	2.6 × 10 ⁵	2.6 × 10 ⁷	6.3 × 10 ⁴	24

^a Matings were performed and quantified as described previously (17). Numbers represent the averages obtained from four independent mating assays.

^b Transfer frequency is calculated as the ratio of transconjugants to donors expressed as a percentage.

pSB24.2 *clt* region can complement the pIJ101 *clt* function, the 142-bp *ClT*⁺ region of pIJ101 present on pGSP263 was replaced with the equivalent 142-bp sequence of pSB24.2 (Fig. 2), which had been PCR amplified with *Pfu* DNA polymerase (Stratagene, La Jolla, Calif.) and relevant primers prior to cloning, and the transfer frequency of the resulting plasmid (pGSP313) was then tested as before. As with pSB24.202, the transfer of pGSP313 from strain TK23.42 also occurred at a frequency (24%) comparable to that of pGSP263.

The pSB24.2 *korB* gene can complement the pIJ101 *korB* function. We also examined whether the pSB24.2 *korB* gene could functionally substitute for its pIJ101 counterpart to override the lethal effects of unregulated *kilB* gene expression in *Streptomyces*. When 100 ng of plasmid pGSP317, a pHYG1 derivative containing the unregulated pIJ101 *kilB* gene, was used to transform *S. lividans* TK23 according to the method of Hopwood et al. (7), a relatively low number of slow-growing transformants were observed after 4 days (i.e., an average of 2.8×10^2 transformants with a diameter of 1 mm or less was obtained in three independent assays). These transformants eventually grew to normal size and were therefore likely to be equivalent to similar transformants obtained previously (12) in which incoming plasmids carrying the unregulated *kilB* gene had subsequently deleted their *kilB* sequences. When TK23 was transformed with an equimolar amount of plasmid pGSP322, which is a derivative of pGSP317 containing the pSB24.2 *korB* gene, an average of 1.0×10^3 faster-growing transformants (i.e., with a diameter of greater than or equal to 2 mm after 4 days) was obtained in three independent assays; such numbers and growth phenotypes were similar to the results seen (i.e., an average of 1.8×10^3 faster-growing transformants) when TK23 was transformed with an equimolar amount of plasmid pGSP321, a pGSP317 derivative containing the pIJ101 *korB* gene.

Results here demonstrating that two important conjugation functions (i.e., *clt* and *korB*) have been conserved between plasmids pIJ101 and pSB24.2 are consistent with the hypothesis that pIJ101 and the parental plasmid pSB24.1 most likely encode the same conjugation system. Given the high degree of overall sequence relatedness between pIJ101 and pSB24.2, plasmids pIJ101 and pSB24.1 (along with its deletion derivative pSB24.2) may also encode highly similar, if not identical, replication systems. The possibility, however, that pIJ101 and pSB24.1 instead represent separate replicons possessing the same conjugation system is intriguing, as we are unaware of any example from other bacterial conjugation systems in which specific *cis*-acting transfer sequences (e.g., *oriT* sequences) (27) encoded by distinct replicons are functionally interchangeable.

The hosts of pIJ101 and pSB24.1 (i.e., *S. lividans* and *S. cyanogenus*, respectively) have 99.8% similarity in their 16S rRNA gene sequences, indicating that they are closely related species (18). Our results are therefore consistent with the notion that conjugative pIJ101-like plasmids may be dispersed throughout at least a portion of the *Streptomyces* genus. Such dispersal has the potential to be even more widespread, however, given the extremely high, albeit limited, degree of relatedness between pIJ101 and pSN22, two plasmids that are derived from species (i.e., *S. lividans* and *S. nigrificans*, respectively) that have only 95.7% homology in their 16S rRNA genes and thus fall into different phylogenetic clusters (18).

Nucleotide sequence accession number. The revised pSB24.2 *korB-clt* sequence determined in this study has been deposited in the GenBank database under accession no. AF133709.

We are very grateful to Alexei Sorokin and Sergey Sineoky, for their assistance in obtaining plasmid pSB24.2, and to Fred A. Rainey for assistance with nucleotide sequencing. We thank Naomi Ward-Rainey, John Battista, and Matthew Ducote for critical reading of the manuscript.

This work was supported by grant MCB-9604879 from the National Science Foundation (to G.S.P.).

REFERENCES

- Altschul, S. F., T. L. Madden, A. A. Schaffer, J. Zhang, Z. Zhang, W. Miller, and D. J. Lipman. 1997. Gapped BLAST and PSI-BLAST: a new generation of protein database search programs. *Nucleic Acids Res.* **25**:3389–3402.
- Bolotin, A. P., A. V. Sorokin, N. N. Aleksandrov, V. N. Danilenko, and Y. I. Kozlov. 1986. Nucleotide sequence of DNA of the actinomycete plasmid pSB24.2. *Dokl. Biochem.* **283**:260–263.
- Brasch, M. A., and S. N. Cohen. 1995. Sequences essential for replication of plasmid pIJ101 in *Streptomyces lividans*. *Plasmid* **33**:191–197.
- Deng, Z., T. Kieser, and D. A. Hopwood. 1988. "Strong incompatibility" between derivatives of the *Streptomyces* multi-copy plasmid pIJ101. *Mol. Gen. Genet.* **214**:286–294.
- Fernandez-Gonzalez, C., R. F. Cadenas, M. F. Noirot-Gros, J. F. Martin, and J. A. Gil. 1994. Characterization of a region of plasmid pBL1 of *Brevibacterium lactofermentum* involved in replication via the rolling circle model. *J. Bacteriol.* **176**:3154–3161.
- Gruss, A., and S. D. Ehrlich. 1989. The family of highly interrelated single-stranded deoxyribonucleic acid plasmids. *Microbiol. Rev.* **53**:231–241.
- Hopwood, D. A., M. J. Bibb, K. F. Chater, T. Kieser, C. J. Bruton, H. M. Kieser, D. J. Lydiate, C. P. Smith, J. M. Ward, and H. Schrempf. 1985. Genetic manipulation of *Streptomyces*: a laboratory manual. John Innes Foundation, Norwich, England.
- Hopwood, D. A., and T. Kieser. 1993. Conjugative plasmids of *Streptomyces*, p. 293–311. *In* D. B. Clewell (ed.), *Bacterial conjugation*. Plenum Press, New York, N.Y.
- Ilyina, T. V., and E. V. Koonin. 1992. Conserved sequence motifs in the initiator proteins for rolling circle DNA replication encoded by diverse replicons from eubacteria, eucaryotes and archaebacteria. *Nucleic Acids Res.* **20**:3279–3285.
- Kataoka, M., Y. M. Kiyose, Y. Michisuiji, T. Horiguchi, T. Seiki, and T. Yoshida. 1994. Complete nucleotide sequence of the *Streptomyces nigrificans* plasmid, pSN22: genetic organization and correlation with genetic properties. *Plasmid* **32**:55–69.
- Kendall, K. J., and S. N. Cohen. 1988. Complete nucleotide sequence of the *Streptomyces lividans* plasmid pIJ101 and correlation of the sequence with genetic properties. *J. Bacteriol.* **170**:4634–4651.
- Kendall, K. J., and S. N. Cohen. 1987. Plasmid transfer in *Streptomyces lividans*: identification of a *kil-kor* system associated with the transfer region of pIJ101. *J. Bacteriol.* **169**:4177–4183.
- Khan, S. A. 1997. Rolling-circle replication of bacterial plasmids. *Microbiol. Mol. Biol. Rev.* **61**:442–455.
- Kieser, T., D. A. Hopwood, H. M. Wright, and C. J. Thompson. 1982. pIJ101, a multi-copy broad host-range *Streptomyces* plasmid: functional analysis and development of DNA cloning vectors. *Mol. Gen. Genet.* **185**:223–238.
- Pabo, C. O., and R. T. Sauer. 1984. Protein-DNA recognition. *Annu. Rev. Biochem.* **53**:293–321.
- Pettis, G. S., and S. N. Cohen. 1996. Plasmid transfer and expression of the transfer (*tra*) gene product of plasmid pIJ101 are temporally-regulated during the *Streptomyces lividans* life cycle. *Mol. Microbiol.* **19**:1127–1135.
- Pettis, G. S., and S. N. Cohen. 1994. Transfer of the pIJ101 plasmid in *Streptomyces lividans* requires a *cis*-acting function dispensable for chromosomal gene transfer. *Mol. Microbiol.* **13**:955–964.
- Rainey, F. A. Personal communication.
- Servín-González, L. 1996. Identification and properties of a novel *clt* locus in the *Streptomyces phaeochromogenes* plasmid pJV1. *J. Bacteriol.* **178**:4323–4326.
- Servín-González, L. 1993. Relationship between the replication functions of *Streptomyces* plasmids pJV1 and pIJ101. *Plasmid* **30**:131–140.
- Stein, D. S., and S. N. Cohen. 1990. Mutational and functional analysis of the *korA* and *korB* gene products of *Streptomyces* plasmid pIJ101. *Mol. Gen. Genet.* **222**:337–344.
- Stein, D. S., K. J. Kendall, and S. N. Cohen. 1989. Identification and analysis of transcriptional regulatory signals for the *kil* and *kor* loci of *Streptomyces* plasmid pIJ101. *J. Bacteriol.* **171**:5768–5775.
- Suzuki, I., M. Kataoka, T. Seki, and T. Yoshida. 1997. Three single-strand origins located on both strands of the *Streptomyces* rolling circle plasmid pSN22. *Plasmid* **37**:51–64.
- Tai, J. T.-N., and S. N. Cohen. 1993. The active form of the KorB protein encoded by the *Streptomyces* plasmid pIJ101 is a processed product that binds differentially to the two promoters it regulates. *J. Bacteriol.* **175**:6996–7005.
- Tai, J. T.-N., and S. N. Cohen. 1994. Mutations that affect regulation of the

- korB* gene of *Streptomyces lividans* plasmid pIJ101 alter plasmid transmission. Mol. Microbiol. **12**:31–39.
26. **Westrich, L., S. Domann, B. Faust, D. Bedford, D. A. Hopwood, and A. Bechthold.** 1999. Cloning and characterization of a gene cluster from *Streptomyces cyanogenus* S136 probably involved in landomycin biosynthesis. FEMS Microbiol. Lett. **170**:381–387.
27. **Wilkins, B., and E. Lanka.** 1993. DNA processing and replication during plasmid transfer between Gram-negative bacteria, p. 105–136. In D. B. Clewell (ed.), Bacterial conjugation. Plenum Press, New York, N.Y.
28. **Zaman, S., L. Radnedge, H. Richards, and J. M. Ward.** 1993. Analysis of the site for second-strand initiation during replication of the *Streptomyces* plasmid pIJ101. J. Gen. Microbiol. **139**:669–676.
29. **Zaman, S., H. Richards, and J. Ward.** 1993. Identification of the minimal replicon of the streptomycete plasmid pIJ101. Plasmid **29**:57–62.