

1-1-2006

## Sequence variation in the coding region of the melanocortin-1 receptor gene (MC1R) is not associated with plumage variation in the blue-crowned manakin (*Lepidothrix coronata*)

Z. A. Cheviron  
*Louisiana State University*

Shannon J. Hackett  
*Field Museum of Natural History*

Robb T. Brumfield  
*Louisiana State University*

Follow this and additional works at: [https://digitalcommons.lsu.edu/biosci\\_pubs](https://digitalcommons.lsu.edu/biosci_pubs)

---

### Recommended Citation

Cheviron, Z., Hackett, S., & Brumfield, R. (2006). Sequence variation in the coding region of the melanocortin-1 receptor gene (MC1R) is not associated with plumage variation in the blue-crowned manakin (*Lepidothrix coronata*). *Proceedings of the Royal Society B: Biological Sciences*, 273 (1594), 1613-1618. <https://doi.org/10.1098/rspb.2006.3499>

This Article is brought to you for free and open access by the Department of Biological Sciences at LSU Digital Commons. It has been accepted for inclusion in Faculty Publications by an authorized administrator of LSU Digital Commons. For more information, please contact [ir@lsu.edu](mailto:ir@lsu.edu).

# Sequence variation in the coding region of the melanocortin-1 receptor gene (*MC1R*) is not associated with plumage variation in the blue-crowned manakin (*Lepidothrix coronata*)

Z. A. Cheviron<sup>1,2,\*</sup>, Shannon J. Hackett<sup>3</sup> and Robb T. Brumfield<sup>1</sup>

<sup>1</sup>Louisiana State University Museum of Natural Science, 119 Foster Hall, Baton Rouge, LA 70803, USA

<sup>2</sup>Department of Biological Sciences, Louisiana State University, Baton Rouge, LA 70803, USA

<sup>3</sup>Department of Zoology, Field Museum of Natural History, 1400 South Lake Shore Drive, Chicago, IL 60605, USA

Avian plumage traits are the targets of both natural and sexual selection. Consequently, genetic changes resulting in plumage variation among closely related taxa might represent important evolutionary events. The molecular basis of such differences, however, is unknown in most cases. Sequence variation in the melanocortin-1 receptor gene (*MC1R*) is associated with melanistic phenotypes in many vertebrate taxa, including several avian species. The blue-crowned manakin (*Lepidothrix coronata*), a widespread, sexually dichromatic passerine, exhibits striking geographic variation in male plumage colour across its range in southern Central America and western Amazonia. Northern males are black with brilliant blue crowns whereas southern males are green with lighter blue crowns. We sequenced 810 bp of the *MC1R* coding region in 23 individuals spanning the range of male plumage variation. The only variable sites we detected among *L. coronata* sequences were four synonymous substitutions, none of which were strictly associated with either plumage type. Similarly, comparative analyses showed that *L. coronata* sequences were monomorphic at the three amino acid sites hypothesized to be functionally important in other birds. These results demonstrate that genes other than *MC1R* underlie melanic plumage polymorphism in blue-crowned manakins.

**Keywords:** ecological genetics; *Lepidothrix*; melanocortin-1 receptor; melanism; plumage polymorphism; sexual selection

## 1. INTRODUCTION

The diversity of avian plumage colours and patterns has fascinated naturalists for centuries. As the targets of both natural and sexual selection, plumage traits can have direct effects on individual fitness, and thus, evolutionary changes in plumage traits may represent important mechanisms in avian speciation (Price 1998). Plumage traits often vary considerably among closely related species, suggesting rapid evolution of plumage differences, which is consistent with their importance as reproductive isolating mechanisms (Omland & Lanyon 2000). Despite the importance of plumage traits in avian speciation, the molecular genetic and developmental pathways responsible for plumage differences among closely related species are entirely unknown in almost all cases.

Elucidating the molecular genetic control of adaptive phenotypic traits, such as many avian plumage characters, is a central goal of evolutionary biology. Only recently, however, it has become possible to explore the basis of such traits in non-model organisms. In this regard, the melanocortin-1 receptor gene (*MC1R*) is of great interest due to its role in melanistic plumage and pelage patterns (Kijas *et al.* 1998; Theron *et al.* 2001; Andersson 2003; Eizirik *et al.* 2003; Hoekstra & Nachman 2003; Doucet *et al.* 2004; Mundy *et al.* 2004; Hoekstra *et al.* 2004;

Rosenblum *et al.* 2004; Mundy 2005). Variation at *MC1R* underlies melanic phenotypes in avian, mammalian and reptilian taxa. In birds, *MC1R* variation is associated with striking melanic plumage polymorphism in bananaquits (*Coereba flaveola*; Theron *et al.* 2001), snow geese (*Anser caerulescens*) (Mundy *et al.* 2004), Arctic skuas (*Stercorarius parasiticus*; Mundy *et al.* 2004), white-winged fairywrens (*Malurus leucopterus*; Doucet *et al.* 2004) and domestic chickens (*Gallus gallus*; Takeuchi *et al.* 1996; Andersson 2003). The taxonomic breadth of this survey suggests that *MC1R* variation may underlie plumage polymorphism in many avian taxa. Indeed, we are aware of only a single published case in which *MC1R* variation was not associated with plumage polymorphism. MacDougall-Shackleton *et al.* (2003) showed that *MC1R* variation does not underlie unmelanized plumage patterns in *Phylloscopus* warblers. This result, however, may have been expected since plumage patterns of many *Phylloscopus* warblers involve complex patterns of unmelanized wing bars, crown stripes and rump patches on a greenish grey background (Mundy 2005), rather than the typical pattern of polymorphism consisting of one predominately dark phenotype and one light phenotype exhibited by bananaquits, skuas, snow geese and other vertebrate taxa (e.g. rock pocket mice (*Chaetodipus intermedius*)—Hoekstra & Nachman 2003). Here, we show that *MC1R* variation does not underlie plumage polymorphism in the blue-crowned manakin (*Lepidothrix*

\* Author for correspondence (zchev1@lsu.edu).

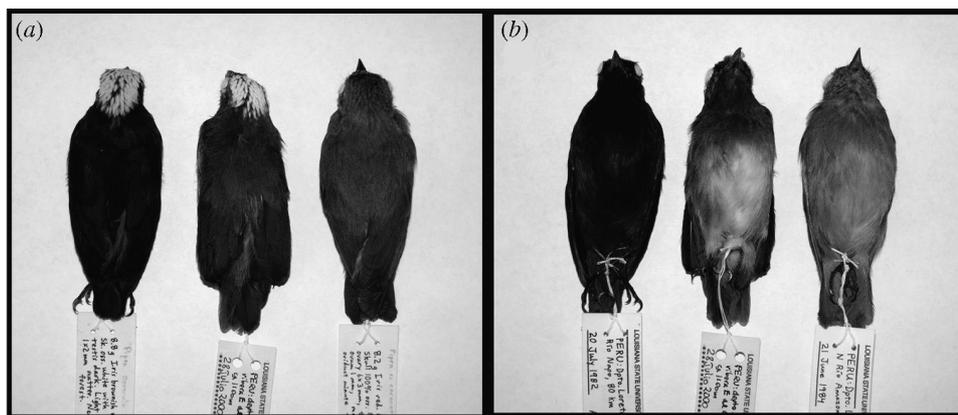


Figure 1. Variation in *L. coronata* male plumage colour. From left to right in both frames is a black male (LSUMZ 110508), a green male (LSUMZ 133193), and a female (LSUMZ 120035). The male specimens correspond, respectively, to tissue samples B-3105 and B-40543 sequenced in this study, and have the same *MC1R* genotype (see table 2). (a) Dorsal view. (b) Ventral view.

*coronata*), a species that exhibits a light–dark geographic pattern of plumage variation.

*Lepidothrix coronata* is a widespread neotropical suboscine passerine, with a striking degree of geographic variation in male plumage colour across its range (Snow 2004; figure 1). In the northern portion of its range (Costa Rica south to northern Peru) males are predominately black with a blue crown patch. In the southern portion of the range (southern Peru to northern Bolivia), males are predominately green with a blue crown patch (figure 1). A broad zone of intergradation between these two plumage groups exists in east-central Peru and western Brazil (Haffer 1970). In this zone, male plumage is generally greenish black, with the degree of green or black plumage in introgressed birds depending on the proximity to the parental populations (Haffer 1970). Males on the southern end of the zone of intergradation are greener than those on the northern end. Throughout the entire range, females are entirely green and lack the blue crown patch.

Male plumage is not concordant with mtDNA phylogeographic patterns (Cheviron *et al.* 2005). Phylogeographic analyses revealed a monophyletic mtDNA haplotype clade in central Peru that encompassed a large proportion of the spectrum of plumage variation, consisting of individuals with pure black, pure green and intermediate plumage (Cheviron *et al.* 2005). Conversely, populations on the north bank of the Amazon river are identical in plumage colour to those on the south bank of the Amazon, yet despite this similarity in plumage, individuals from these populations fall into relatively distantly related haplotype clades (Cheviron *et al.* 2005). These results suggest that the microevolutionary forces shaping geographic variation in male plumage may differ from those shaping phylogeographic mtDNA patterns. Since *L. coronata* is a polygynous, lekking species (Snow 2004), male plumage traits, and the genes underlying them, may be under strong sexual selection, possibly explaining incongruence between mtDNA phylogeographic patterns and geographic variation in male plumage (Brumfield *et al.* 2001).

In this study, we examined sequence variation in the coding region of *MC1R* to test the hypothesis that *MC1R* variation is associated with variation in male plumage colour in *L. coronata*.

## 2. MATERIAL AND METHODS

### (a) Taxon sampling

We sampled 23 individuals from across the range of *L. coronata* (table 1), with most samples coming from the central Amazonian portion of the range that encompasses the zone of intergradation between the green and black plumage groups. North and south of this zone of intergradation, male plumage colour is relatively invariant. We included representatives of all the phylogeographic mtDNA clades identified by Cheviron *et al.* (2005), as well as all of the major plumage types. To determine polarity of substitutions, we sequenced *MC1R* from the congeneric white-crowned manakin (*L. serena*).

### (b) Amplification and sequencing of *MC1R*

Total genomic DNA was extracted from approximately 25 mg of pectoral muscle tissue using QIAGEN DNeasy extraction kits (QIAGEN, Valencia, California). We amplified an 810 bp fragment of the avian *MC1R* gene using the following primers modified from Mundy *et al.* (2004): lcorMSHR9—5'CTGGCTCCGGAAGGCRTAGAT 3' and lcorMSHR72—5'AYGCCAGYGAGGGCAACCA3'. This fragment, which corresponds to bases 70–880 of the chicken *MC1R* gene (Kerje *et al.* 2003), includes all sites previously shown to be associated with melanic phenotypes in birds. We performed polymerase chain reactions (PCRs) in 25 µl volumes, with 0.1 µl AmpliTaq DNA polymerase (Applied Biosystems, Foster City, California), 2.5 µl 10X Tris buffer with MgCl<sub>2</sub> (Applied Biosystems), 1.5 µl dNTPs (each dNTP 150 µM), 1.5 µl (600 nM) of each primer and 2.5 µl (approx. 50 ng) of template DNA using a PTC-200 thermal cycler (MJ Research, Waltham, Massachusetts). PCR thermocycling conditions were: (i) an initial denaturing step of 94 °C for 2 min; (ii) 35 cycles of the following: 30 s at 94 °C, 45 s at 62.5 °C and 90 s at 72 °C; and (iii) a 5 min extension step at 72 °C. After purifying amplicons with PEG precipitation, we performed sequencing reactions using an ABI Prism cycle sequencing kit (Applied Biosystems) with the PCR primers and the internal sequencing primers: lcorMSHR73—5'SGCRTAGAAGATGGTGTATGTAGC 3' and lcorMSHR74—5'GTBACCGCTACATCACCAT 3'. We purified sequencing products using AutoSeq G-50 (Amersham, Uppsala, Sweden) spin columns and visualized them using an ABI 3100 Genetic Analyzer. All sequence data

Table 1. Sampling localities and voucher numbers of specimens used to examine MC1R sequence variation in *L. coronata*. (Geographic coordinates were obtained either from specimen tags or museum catalogues.)

locality	species	tissue source	tissue voucher numbers
Costa Rica: Puntarenas; Rio Copey, 4 km E Jaco, 9.62°, -84.63°	<i>L. coronata</i>	LSUMZ	B-16077
Panama: Pamama; E Panama Canal, 9.33°, -79.92°	<i>L. coronata</i>	LSUMZ	B-28461
Venezuela: Amazonas; San Carlos de Rio Negro, -2.08°, -66.48°	<i>L. coronata</i>	AMNH	RWD17053
Peru: San Martin; 20 km NE Tarapoto on road to Yurimaguas, -6.41°, -75.66°	<i>L. coronata</i>	LSUMZ	B-5394
	<i>L. coronata</i>	LSUMZ	B-5487
Peru: Loreto; 1.5 km S Libertad, S bank of Rio Napo, 80 km N Iquitos, -3.03°, -72.75°	<i>L. coronata</i>	LSUMZ	B-3105
Peru: Loreto; S bank of Rio Amazonas, ca 10 km SSW mouth of Rio Napo, -3.42°, -72.58°	<i>L. coronata</i>	LSUMZ	B-4657
Peru: Loreto; lower Rio Napo region, ca 90 km N Iquitos, -3.91°, -73.08°	<i>L. coronata</i>	LSUMZ	B-4545
Peru: Loreto; 86 km SE Juanjui, -7.58°, -75.93°	<i>L. coronata</i>	LSUMZ	B-40543
Peru: Ucayali; SE slope of Cerro Tahuayo, -8.13°, -74.04°	<i>L. coronata</i>	LSUMZ	B-10492
Peru: Loreto; 79 km WNW Contamana, -7.13°, -75.68°	<i>L. coronata</i>	LSUMZ	B-27578
	<i>L. coronata</i>	LSUMZ	B-27832
Peru: Madre de Dios; Moskitania, 13.4 km NNW Atalaya, -13.82°, -71.63°	<i>L. coronata</i>	FMNH	B-433689
	<i>L. coronata</i>	FMNH	B-433691
	<i>L. coronata</i>	FMNH	B-433692
	<i>L. coronata</i>	FMNH	B-433693
	<i>L. coronata</i>	FMNH	B-433694
Brazil: Rondônia; ca 50 km NW Jaciparana, w bank Rio Maderia, -9.25°, -64.40°	<i>L. coronata</i>	LSUMZ	B-31333
Bolivia: La Paz Department; Rio Beni, ca 20 km by river N Puerto Linares, -12.56°, -67.20°	<i>L. coronata</i>	LSUMZ	B-901
Bolivia: Pando Department; Nicolás Suarez, 12 km by road S Cobija, -11.18°, -69.03°	<i>L. coronata</i>	LSUMZ	B-9125
	<i>L. coronata</i>	LSUMZ	B-9177
	<i>L. coronata</i>	LSUMZ	B-9269
	<i>L. coronata</i>	LSUMZ	B-9306
Brazil: Amazonas Department; ca 80 km N Manaus	<i>L. serena</i>	LSUMZ	B-20306

generated in this study were deposited in GenBank (accession numbers DQ388308–DQ388331).

### (c) Genetic analyses

We aligned sequences from both strands by eye using SEQUENCHER, v. 4.1 (Genecodes, Ann Arbor, Michigan). In the chromatograms of three individuals (table 2), we noted double peaks at single sites that were approximately half the height of neighbouring peaks. We coded individuals as heterozygous if such double peaks were noted in both strands (MacDougall-Shackleton *et al.* 2003). We aligned *Lepidothrix* MC1R sequences with *G. gallus* (GenBank—AY220305), *C. flaveola* (GenBank—AF362605 and AF362598), *A. caerulescens* (AY521209 and AY521182) and *S. parasiticus* (GenBank—AY521214 and AY521217) MC1R sequences to confirm identity of the amplicons and to translate nucleotide sequences to amino acid sequences. Alignment with these other avian sequences allowed us to examine sequences for indels and stop codons indicative of pseudogenes, neither of which was observed. This comparative analysis also allowed for the examination of variation at nucleotide sites that were previously shown to be associated with melanistic plumage in other birds.

We tested for evidence of selection at MC1R by calculating Tajima's *D* statistic (Tajima 1989) using the program DNASP 3.53 (Rozas & Rozas 1999).

## 3. RESULTS

### (a) MC1R sequence variation and its association with male plumage colour

A total of 810 bp of the MC1R gene was sequenced for 23 *L. coronata* individuals corresponding to nucleotide positions 70–880 of the *G. gallus* MC1R gene (Kerje *et al.* 2003). Four sites were variable within *L. coronata*, resulting in a total of four MC1R alleles, none of which appeared to be recombinants based on the four gamete test (Hudson & Kaplan 1985; table 2). All the substitutions were synonymous and none showed any association to the two plumage colour groups (table 2). Thus, we found no evidence that sequence variation at MC1R is associated with plumage colour variation in *L. coronata*.

Sequence variation at MC1R was largely consistent with phylogeographic patterns inferred from mtDNA sequence variation (Cheviron *et al.* 2005). Two synapomorphic nucleotide sites (78 and 204) support the split between the *cis*-Andean (east of the Andes) and *trans*-Andean (west of the Andes) lineages, which is the basal split inferred from mtDNA (Cheviron *et al.* 2005). One site (411) varies across the plumage zone of intergradation, and the transition is largely concordant with the southern edge of the zone of intergradation. Variation at this site, however, is not associated with plumage colour as we sampled pure green individuals that were homozygous for each allele (table 2).

Table 2. *MC1R* sequence polymorphism data for 23 *L. coronata* individuals. Asterisks indicate agreement with the consensus sequence. All variation among the four *L. coronata* alleles is synonymous with respect to the amino acid sequence. Heterozygous sites are denoted with standard IUPAC one-letter codes (W=A/T). Genotype refers to allelic combinations. Sites highlighted in grey are putatively functional nucleotide sites in other avian taxa. Substitutions at site 253 are associated with plumage polymorphism in snow geese (*A. caerulescens*) (Mundy *et al.* 2004). Substitutions at site 274 are associated with plumage polymorphism in bananaquits (*Coereba flaveola*) (Theron *et al.* 2001). Substitutions at site 689 are associated with plumage polymorphism in Arctic skuas (*S. parasiticus*; Mundy *et al.* 2004). All *L. coronata* individuals are monomorphic at these three sites. Nucleotide sites are numbered with reference to the chicken *MC1R* sequence. *Lepidothrix serena* is included as an outgroup.

voucher number	nucleotide site										plumage	genotype	genbank accession number
	7	9	0	2	2	3	4	6	6	8			
consensus	C	G	C	G	G	C	A	C	A	A			
B-28461	T	*	T	*	*	*	*	*	*	*	black	1/1	DQ388308
B-16077	T	*	T	*	*	*	*	*	*	*	black	1/1	DQ388309
RWD17053	*	*	*	*	*	*	*	*	*	*	black	2/2	DQ388310
B-3105	*	*	*	*	*	*	*	*	*	*	black	2/2	DQ388313
B-4657	*	*	*	*	*	*	*	*	*	*	black	2/2	DQ388314
B-4545	*	*	*	*	*	*	*	*	*	*	black	2/2	DQ388315
B-10492	*	*	*	*	*	*	*	*	*	*	intermediate	2/2	DQ388316
B-27578	*	*	*	*	*	*	*	*	*	*	intermediate	2/2	DQ388318
B-27832	*	*	*	*	*	*	*	*	*	*	intermediate	2/2	DQ388319
B-5394	*	*	*	*	*	*	*	*	*	*	green	2/2	DQ388311
B-5487	*	*	*	*	*	*	*	*	*	*	green	2/2	DQ388312
B-40543	*	*	*	*	*	*	*	*	*	*	green	2/2	DQ388317
B-901	*	*	*	*	*	*	*	*	*	*	green	2/2	DQ388320
B-433693	*	*	*	*	*	*	*	*	*	*	green	2/2	DQ388321
B-433692	*	*	*	*	*	*	*	*	*	*	green	2/2	DQ388322
B-433689	*	*	*	*	*	*	*	*	*	*	green	2/2	DQ388323
B-433691	*	*	*	*	*	*	W	*	*	*	green	2/3	DQ388324
B-433694	*	*	*	*	*	*	W	*	*	*	green	2/3	DQ388325
B-9269	*	*	*	*	*	*	W	*	*	*	green	2/3	DQ388326
B-9306	*	*	*	*	*	T	T	*	*	*	green	4/4	DQ388327
B-31333	*	*	*	*	*	*	T	*	*	*	green	3/3	DQ388328
B-9125	*	*	*	*	*	*	T	*	*	*	green	3/3	DQ388329
B-9177	*	*	*	*	*	*	T	*	*	*	green	3/3	DQ388330
<i>L. serena</i>	T	A	*	*	*	*	*	T	*	*	n.a	n.a	DQ388331

#### (b) Nature of selection at *MC1R* in *L. coronata*

Values of Tajima's *D* did not differ significantly from zero ( $-0.4168$ ,  $p > 0.10$ ), thus we failed to reject a null hypothesis of neutral variation. However, we noted that since only a few variable sites were observed, our statistical power to detect selection was low.

#### 4. DISCUSSION

Despite its apparent role in plumage and pelage melanism in several other vertebrate taxa (reviewed in Mundy 2005), we found no evidence that sequence variation at *MC1R* is associated with male plumage variation in *L. coronata*. We did not detect any non-synonymous differences among *MC1R* sequences of green and black plumaged birds, and the synonymous variation at *MC1R* we did uncover showed no association with plumage colour (table 2). In snow geese (*A. caerulescens*) and bananaquits (*C. flaveola*), single non-synonymous substitutions Val<sup>85</sup>→Met<sup>85</sup> and Glu<sup>92</sup>→Lys<sup>92</sup>, respectively, are associated with melanistic plumage (Theron *et al.* 2001; Mundy *et al.* 2004). All of the blue-crowned manakins sampled here were monomorphic for the amino acids associated with non-melanistic plumage in these taxa (Val and Glu; table 2), demonstrating that genes

other than *MC1R* underlie variation in plumage melanism in *L. coronata*. The pattern was similar at a third putatively functional *MC1R* site. In Arctic skuas, a non-synonymous substitution (Arg<sup>230</sup>→His<sup>230</sup>) is associated with melanistic plumage (Mundy *et al.* 2004). Again, all of the manakins screened here were monomorphic at this site, but in this case, the amino acid associated with melanistic plumage, histidine, was fixed. This result is particularly noteworthy because Mundy *et al.* (2004) suggested the Arg<sup>230</sup>→His<sup>230</sup> substitution was significant because a histidine at the homologous site in rock pocket mice (*Chaetodipus intermedius*) is also associated with melanistic pelage (Hoekstra & Nachman 2003; Nachman *et al.* 2003). These data show clearly that a histidine at this amino acid position does not confer melanistic plumage in all avian genetic backgrounds.

To our knowledge, this is only the second documented case of a lack of a relationship between melanistic plumage and *MC1R* sequence variation. These data, in conjunction with the negative results reported in *Phylloscopus* warblers (MacDougall-Shackleton *et al.* 2003), demonstrate that in some cases, genes other than *MC1R* underlie melanistic plumage polymorphism, suggesting that variation exists in the genetic basis of melanistic plumage traits. Indeed, such

variation has already been demonstrated in rock pocket mice, within which melanic pelage is correlated with *MC1R* variation in some populations, but not in others (Hoekstra & Nachman 2003).

Since we did not sequence the first 23 and last 20 codons of *MC1R*, we cannot rule out entirely the presence of functionally important variation in these regions. However, the gene portion sequenced here contained all of the sites previously shown to be functionally important in birds, all of which lie in trans-membrane or cytoplasmic domains (Theron *et al.* 2001; Kerje *et al.* 2003; Mundy *et al.* 2004; Mundy 2005). The unsequenced regions are not parts of these domains, and we are unaware of any studies that have reported functionally important sites in these unsequenced regions in any taxon. Thus, it is unlikely that functionally important variation would exist in these regions.

First-year male *L. coronata* are green, resembling females, and probably acquire their definitive adult plumage during their second year. This ontogenetic pattern suggests a regulatory gene may influence adult male plumage polymorphism. Moreover, the diversity of intermediate plumage types within the zone of intergradation suggests that male plumage colour is a polygenic trait. Together, these observations indicate the potential roles of molecular genetic and development pathways in the control of plumage trait variation in this taxon. Many avian species exhibit similar ontogenetic patterns with males developing definitive plumage later in life. Since adult plumage is the target of sexual selection in many species, further studies of taxa with delayed plumage maturation are needed.

Over 100 loci are known to influence levels of pigmentation in vertebrates (Smyth 1990; Bennett & Lamoreux 2003). Future studies exploring other candidate genes, especially those with regulatory functions are likely to provide great insight into our understanding of the evolution of avian plumage colour.

We wish to thank the following institutions for loaning tissue samples and specimens used in this study: the American Museum of Natural History, The Field Museum of Natural History, the Louisiana State University Museum of Natural Science. Constructive criticisms from Matthew D. Carling, Ellen D. Ketterson, and three anonymous reviewers greatly improved the manuscript. Funding for this project was provided by the Frank M. Champman Memorial Fund of the American Museum of Natural History (Z.A.C.) and NSF grants DEB-0332093 and DBI-0400797 (R.T.B.).

## REFERENCES

- Andersson, L. 2003 Melanocortin variants with phenotypic effects in horse, pig, and chicken. *Ann. N. Y. Acad. Sci.* **994**, 313–318.
- Bennett, D. C. & Lamoreux, M. L. 2003 The color loci of mice—a genetic century. *Pigment Cell Res.* **16**, 333–344. (doi:10.1034/j.1600-0749.2003.00067.x)
- Brumfield, R. T., Jernigan, R. W., McDonald, D. B. & Braun, M. J. 2001 Evolutionary implications of divergent clines in an avian (*Manacus*; Aves) hybrid zone. *Evolution* **55**, 2070–2087.
- Cheviron, Z. A., Hackett, S. J. & Capparella, A. P. 2005 Complex evolutionary history of a Neotropical lowland forest bird (*Lepidothrix coronata*) and its implications for historical hypotheses of the origin of Neotropical avian diversity. *Mol. Phylogenet. Evol.* **36**, 336–357.
- Doucet, S. M., Shawkey, M. D., Rathburn, M. K., Mays, H. L. & Montgomerie, R. 2004 Concordant evolution of plumage colour, feather microstructure and a melanocortin receptor gene between mainland and island populations of a fairy-wren. *Proc. R. Soc. B* **271**, 1663–1670. (doi:10.1098/rspb.2004.2779)
- Eizirik, E., Yuhki, N., Johnson, W. E., Menotti-Raymond, M., Hannah, S. S. & O'Brien, S. J. 2003 Molecular genetics and evolution of melanism in the cat family. *Curr. Biol.* **13**, 448–453. (doi:10.1016/S0960-9822(03)00128-3)
- Haffer, J. 1970 Speciation in some Amazonian forest birds. *J. Ornithol.* **111**, 285–331. (doi:10.1007/BF01653396)
- Hoekstra, H. E. & Nachman, M. W. 2003 Different genes underlie adaptive melanism in different populations of rock pocket mice. *Mol. Ecol.* **12**, 1185–1194. (doi:10.1046/j.1365-294X.2003.01788.x)
- Hoekstra, H. E., Drumm, K. E. & Nachman, M. W. 2004 Ecological genetics of adaptive color polymorphism in pocket mice: geographic variation in selected and neutral genes. *Evolution* **58**, 1329–1341.
- Hudson, R. R. & Kaplan, N. L. 1985 Statistical properties of the number of recombination events in the history of a sample of DNA sequences. *Genetics* **111**, 147–164.
- Kerje, S., Lind, J., Schutz, K., Jensen, P. & Andersson, L. 2003 Melanocortin 1-receptor (*MC1R*) mutations are associated with plumage colour in chicken. *Anim. Genet.* **34**, 241–248. (doi:10.1046/j.1365-2052.2003.00991.x)
- Kijas, J. M. H., Wales, R., Tornsten, A., Chardon, P., Moller, M. & Andersson, L. 1998 Melanocortin receptor 1 (*MC1R*) mutations and coat color in pigs. *Genetics* **150**, 1177–1185.
- MacDougall-Shackleton, E. A., Blanchard, L. & Gibbs, H. L. 2003 Unmelanized plumage patterns in old world leaf warblers do not correspond to sequence variation at the melanocortin-1 receptor locus (*MC1R*). *Mol. Biol. Evol.* **20**, 1675–1681. (doi:10.1093/molbev/msg186)
- Mundy, N. I. 2005 A window on the genetics of evolution: *MC1R* and plumage colouration in birds. *Proc. R. Soc. B* **272**, 1633–1640. (doi:10.1098/rspb.2005.3107)
- Mundy, N. I., Badcock, N. S., Hart, T., Scribner, K., Janssen, K. & Nadeau, N. J. 2004 Conserved genetic basis of a quantitative plumage trait involved in mate choice. *Science* **303**, 1870–1873. (doi:10.1126/science.1093834)
- Nachman, M. W., Hoekstra, H. E. & D'Agostino, S. L. 2003 The genetic basis of adaptive melanism in pocket mice. *Proc. Natl Acad. Sci.* **100**, 5268–5273. (doi:10.1073/pnas.0431157100)
- Omland, K. E. & Lanyon, S. E. 2000 Reconstructing plumage evolution in orioles (*Icterus*): repeated convergence and reversal in patterns. *Evolution* **54**, 2119–2133.
- Price, T. D. 1998 Sexual selection and natural selection in bird speciation. *Phil. Trans. R. Soc. B* **353**, 251–260. (doi:10.1098/rstb.1998.0207)
- Rosenblum, E. B., Hoekstra, H. E. & Nachman, M. W. 2004 Adaptive reptile color variation and the evolution of the *MC1R* gene. *Evolution* **58**, 1794–1808.
- Rozas, J. & Rozas, R. 1999 DnaSp version 3: an integrated program for molecular population genetics and molecular evolution analysis. *Bioinformatics* **15**, 174–175.
- Smyth, J. R. 1990 Genetics of plumage, skin, and pigmentation in chickens. In *Poultry breeding and genetics* (ed. R. D. Crawford). New York, NY: Elsevier Science.
- Snow, D. 2004 Family Pipridae (manakins). In *Handbook of Birds of the World: contigas to pipits and wagtails*, vol. 9 (ed. J. del Hoyo, A. Elliot & D. Christie), pp. 110–169. Barcelona, Spain: Lynx Edicions.

- Tajima, F. 1989 Statistical method for testing the neutral mutation hypothesis by DNA polymorphism. *Genetics* **123**, 585–595.
- Takeuchi, S., Suzuki, H., Yabuuchi, M. & Takahashi, S. 1996 A possible involvement of the melanocortin-1 receptor in regulating feather pigmentation in the chicken. *Biochim. Biophys. Acta* **1308**, 164–168.
- Theron, E., Hawkins, K., Bermingham, E., Ricklefs, R. E. & Mundy, N. I. 2001 The molecular basis of an avian plumage polymorphism in the wild: a melanocortin-1-receptor point mutation is perfectly associated with the melanic plumage morph of the bananaquit, *Coereba flaveola*. *Curr. Biol.* **11**, 550–557. (doi:10.1016/S0960-9822(01)00158-0)