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The Effect of Diets Varying in Dietary Cation-Anion Difference Fed in Late Gestation and in Lactation on Sow Productivity

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The effect of diets varying in dietary cation-anion difference fed in late gestation and in lactation on sow productivity

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**THE EFFECT OF DIETS VARYING IN DIETARY CATION-ANION DIFFERENCE FED IN LATE
GESTATION AND IN LACTATION ON SOW PRODUCTIVITY**

A Thesis

Submitted to the Graduate Faculty of the
Louisiana State University and
Agricultural and Mechanical College
in partial fulfillment of the
requirements for the degree of
Master of Science

in

The Interdepartmental Program in
Animal, Dairy, and Poultry Sciences

by
Melanie Roux
B. S., Louisiana State University, 2002
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ABSTRACT

Eighty-six primiparous or multiparous sows and their pigs were used to evaluate the effects of changing dietary cation-anion difference (DCAD) in late gestation and in lactation on sow productivity. Twenty sows were used in Exp. 1, and the dietary treatments consisted of a positive control (PC) corn-soybean meal (C-SBM) diet with four levels of DCAD (140, 99, 75, and 45 mEq/kg). These DCAD's were achieved by four levels of SoyChlor (SC) additive (0, 1.5, 2.5, and 3.5%). In Exp. 2, 66 sows were used and the dietary treatments consisted of the diets providing DCAD of 140 and 45 mEq/kg. The diets were fed from d 107 of gestation to weaning in Exp. 1 and from d 111 of gestation to weaning in Exp. 2. Sows were allotted based on parity and their farrowing date.

Experiment 1 was a preliminary experiment to determine the level of DCAD that reduced urinary pH. Reducing DCAD did not affect sow reproductive responses. Urinary pH was linearly decreased ($P < 0.001$) as DCAD decreased in the diet. Reducing DCAD tended to linearly increase ($P = 0.15$) plasma Ca concentrations.

In Exp. 2, reducing DCAD from 140 to 45 mEq/kg reduced ADFI from d 111 of gestation to d 1 postfarrowing ($P < 0.02$), but ADFI was not affected by DCAD during any other period. Reducing DCAD did not affect reproductive performance of the sows nor litter response variables or plasma Ca. Decreasing DCAD in the diet decreased urinary pH ($P < 0.001$). Twenty-seven sows fed the PC diet and 21 sows fed the reduced DCAD diet during the previous lactation were evaluated during their subsequent farrowing. Sows that were fed the reduced DCAD diet had increased total number of pigs born ($P < 0.08$) and pigs born alive ($P < 0.02$) in the subsequent farrowing. Changing DCAD had little effect on sow and litter response variables in the current lactation, but it decreased urine pH ($P < 0.001$), and increased total number of pigs born ($P < 0.08$) and pigs born alive ($P < 0.02$) in the subsequent farrowing.

CHAPTER 1

INTRODUCTION

Feed additives are ingredients that are fed to animals, usually to improve productivity. Acidifiers are feed additives that contain both cations and anions. Cations are minerals with a positive charge. Anions are minerals with a negative charge. Dietary cation-anion difference (DCAD) is the difference between strong fixed cations and strong fixed anions. The differences in ions are used to determine the relationship between strong cations and anions and to predict whether the diet will elicit an acidic or alkaline response when fed to the animal.

Postparturient paresis (milk fever) is a problem in the dairy industry. Altering DCAD is important in dairy nutrition. SoyChlor (**SC**) is an anionic salt feed additive that contains CaCl_2 and HCl . This anionic salt is commonly fed to dairy cattle to make the diet more acidic, thereby preventing milk fever. Individual cases of milk fever differ between herds, but approximately 5 to 7% of adult dairy cows in the United States are afflicted (Jordan et al., 1993). Cows afflicted with milk fever have reduced feed intake, reduced urination and defecation, and if left untreated, cows become comatose and die (Horst et al., 1997). Milk fever contributes to a severe economic loss because milk production declines in the subsequent lactation (Block, 1984) and it is expensive to treat due to medication and veterinarian visits (Block et al., 1989). Sows usually are not afflicted with milk fever, but urinary tract infections can cause the swine industry to suffer economic loss due to reduced herd health. The purpose of this research was to determine if reducing the dietary cation-anion difference in the diets would affect sow productivity when fed during late gestation and in lactation.

CHAPTER 2

REVIEW OF LITERATURE

General

Anionic salts have an increased amount of Cl^- and S^- in relation to Na^+ and K^+ . These four ions are the most important in determining the effects of diet on systemic cation-anion balance (Oetzel et al., 1991). Calcium chloride is an anionic salt routinely added to dairy feed to change the DCAD. Calcium chloride increases dietary Cl^- without increasing Na^+ or K^+ , thus lowering DCAD. An acidifying diet stimulates the release of cations (especially Ca^{2+}) into the blood in order for the animal to maintain homeostatic blood pH. Incorporating anionic salts into the diet helps to increase Ca^{2+} absorption and bone Ca^{2+} mobilization. Research indicates that feeding anionic salts acidifies the blood, which restores responsiveness to the parathyroid hormone receptor sites (NRC, 2001).

A commonly fed anionic salt is CaCl_2 , which is found in the feed additive SC. SoyChlor is a feed additive that supplies the Cl^- anion in a carrier that has been extracted from soybean meal, which means it is palatable and high in protein. SoyChlor contains high amounts of Ca (3.40%) and chloride (9.49%). Chloride is extremely high in SC because it contains two different sources (CaCl_2 and HCl). Hydrochloric acid is a powerful acidifier, therefore less has to be added to the diet to achieve the same results as a less potent anionic salt (West Central Soy). As early as the 1960's, researchers have observed the importance of manipulating DCAD of feedstuffs to reduce the incidences of postparturient paresis (milk fever) in dairy cows. Milk fever is a metabolic disorder that is triggered by the onset of lactation. After parturition, the cow's Ca stores become imbalanced because the cow's demands for Ca exceed her available resources (Purina Dairy News, 2000). The cow's plasma Ca concentration declines due to Ca being lost to colostrum. Milk fever is more prone to develop when a high DCAD diet is consumed during the last three weeks of pregnancy. Literature suggests that diets high in cations tend to induce milk fever, whereas diets high in anions prevent it (Block et al., 1984; Tucker et al., 1991; Goff et al., 1998; Riond, 2001).

Research shows that reducing DCAD minimizes the risk of milk fever by lowering intestinal pH, which affects the amount of Ca absorbed (Tucker et al., 1991). Goff et al. (1998) conducted a study involving HCl and milk fever prevention. They concluded that cows fed HCl (1.5 eq/d) had increased feed intake before calving, increased plasma Ca concentrations, and decreased urinary pH compared to cows fed the control diet. Other research indicates that anionic salts reduce palatability, thereby reducing feed intake (Yen et al., 1981). Goff et al. (1998) indicated that adding HCl to the diet reduced the risk of milk fever. Two experiments were conducted with cows evaluating the effect of cation-anion balance on milk fever (Block, 1984). In the first study, two diets (+33.05 mEq/kg and -12.85 mEq/kg DM) were fed for two years. They reported no differences in feed intake, but cows fed the cationic diet had 47.4% greater incidence of milk fever, while cows fed the anionic diet had no incidence of milk fever. In the second study, Block et al. (1989) fed four diets (400, 200, 100, and 50 mEq/kg DM) to prepartum cows. They concluded that feed intake was lower in cows fed 200 mEq/kg prepartum, but at parturition, feed intake was similar across all treatments. They concluded that decreasing DCAD below the control diet reduced the risk of milk fever. Plasma Ca concentration was increased in the blood when the DCAD was reduced.

Effect of Dietary Cation-Anion Difference on Plasma Ca Concentrations

During gestation, cows lose Ca in relatively small amounts to the excreta and fetus. These losses are not large enough to activate parathyroid hormone mechanisms in the body to increase Ca concentrations during parturition and lactation. Two hormones, parathyroid hormone and 1,25-dihydroxyvitamin D₃ [(1,25(OH)₂ D₃)] work together in regulating plasma Ca (Horst et al., 1997) concentrations. When plasma Ca concentrations decline, parathyroid hormone and 1,25(OH)₂ D₃ are produced to increase Ca (Horst et al., 1997). Bone is a major contributor to maintaining Ca levels in the blood. If Ca levels are too low, parathyroid hormone stimulates the bone to release Ca that is stored in the lining cells and into the extracellular pool. Before reducing DCAD in dairy cows, nutritionists fed a low Ca diet before calving. This decrease in dietary Ca stimulates parathyroid hormone and 1,25(OH)₂ D₃ production to put the cow in positive Ca balance (NRC, 2001).

After parturition, the cow is at less risk to get milk fever because these Ca homeostatic mechanisms are already active. However, this theory is not practical because a cow needs to eat 20 g/d to meet the Ca requirement (West Central Soy). It is difficult to reduce the Ca in a cows diet.

Effect of Dietary Cation-Anion Difference on Urinary pH

Normal urine pH in dairy cows is approximately 8. Including anions in the feed reduces urine pH to approximately 6. When feeding an anionic diet, testing urine pH is the quickest method to determine if the anions are working properly. Riond (2001) reported that a cow's urinary pH needs to be 6.5 to prevent milk fever. Giesy et al. (1997) conducted an experiment with four diets consisting of 30, 10, -10, and -30 mEq DCAD/100g DM to dry cows. They reported that urine pH and DCAD have a strong relationship; as DCAD decreases, urinary pH decreases.

Effect of Dietary Cation-Anion Difference on Sow Growth and Reproductive Performance

DeRouchey et al. (2003) fed sows DCAD's of 0, 100, 200, 350, and 500 mEq/kg. Changing DCAD did not affect overall ADFI, but increasing DCAD linearly decreased percent survivability in pigs. Increasing DCAD linearly decreased plasma Ca concentrations and increased urinary pH. Dove and Haydon et al. (1994) fed sows DCAD levels of 130.8, 161.2, and 250.8 mEq/kg and reported no differences in feed intake, days to estrus, percent survivability in pigs or litter weight gain regardless of dietary treatments. Patience et al. (1987) fed grower pigs a range (-85 to +341 mEq/kg) of DCAD and reported that growth and feed intake were similar for the positive DCAD, but the negative DCAD diets reduced growth and feed intake. Yen et al. (1981) fed three diets; 1) Control, 2) 4% CaCl_2 + 2.22% $\text{Na}_5\text{P}_3\text{O}_{10}$, and 3) 4% CaCl_2 + 2.03% NaHCO_3 to finisher pigs. Diet two (4% CaCl_2 + 2.22% $\text{Na}_5\text{P}_3\text{O}_{10}$) decreased ADFI which resulted in a decrease in ADG and G:F. Plasma Cl^- concentrations were increased from 100 to 112 mmol/L in pigs fed 4% CaCl_2 + 2.22% $\text{Na}_5\text{P}_3\text{O}_{10}$, which may be a reason for the decrease in feed intake. The increased plasma Cl^- concentrations may cause body fluid to become acidogenic, which may reduce feed intake. Austic et al. (1983) fed grower pigs a range of DCAD (-100, 0, 100, 200, 300, and 500 mEq/kg) and reported no differences in ADG or G:F when these DCAD diets were fed.

Conclusions

Further research is needed before specific recommendations can be made regarding optimal DCAD in rations for swine. Research indicates that feeding anionic diets to prepartum and postpartum dairy cows increases plasma Ca, decreases urinary pH, and does not affect feed intake. More research in swine is needed to determine if reducing DCAD will have positive effects on feed intake and plasma Ca concentrations.

CHAPTER 3

THE EFFECT OF DIETS VARYING IN DIETARY CATION-ANION DIFFERENCE FED IN LATE GESTATION AND IN LACTATION ON SOW PRODUCTIVITY

Introduction

Increasing sow and litter performance during lactation is vital to the swine industry (DeRouchey et al., 2003). Sow litter size has increased during the last decade (Agric. Ltd, 2003). This increase in litter size has increased the demands for milk production. Unlike dairy cows, milk fever is not a major problem in the swine industry. However, during late gestation and lactation, sows can become constipated, which causes difficulties in farrowing. Excess constipation can cause reduced feed intake and this leads to a reduction in milk production, which decreases litter growth. Swine nutritionists are evaluating altering DCAD in sow lactation diets (DeRouchey et al., 1998). Sows are prone to urinary tract infections, and anionic salts are increasingly being fed to sows to reduce urinary pH, which may in turn decrease urinary tract infections (DeRouchey et al., 2003). Calcium chloride (CaCl_2), potassium chloride (KCl), and magnesium sulfate (MgSO_4) are mineral compounds that can easily be added to sow diets to decrease urine pH. Research shows that by incorporating anionic salts into the diet of prepartum dairy cows, urinary pH decreases, Ca metabolism is improved, and milk fever is prevented. However, minimal research has been conducted on DCAD and its effects on sows during lactation and on reproductive performance. Therefore, the objective of this experiment was to determine the effects of decreasing DCAD on sow productivity.

Materials and Methods

General

The Louisiana State University (**LSU**) Agricultural Center Animal Care and Use Committee approved all methods used in these experiments. Primiparous and multiparous Yorkshire and crossbred (Yorkshire x Landrace or Yorkshire x Duroc) sows from the LSU Agricultural Center Swine Unit were allotted to dietary treatments on d 107 of gestation for Exp.1 and d 111 of gestation for Exp. 2.

Before starting dietary treatments, all sows were fed a typical C-SBM gestation diet that met or exceeded the nutrient requirements (NRC, 1998) of gestating sows. The sows were allotted to their respective treatments within each group based on parity and the date of d 107 of gestation for Exp.1 and d 111 of gestation for Exp.2.

Diets fed during both experiments were formulated on a total AA basis from analyzed AA values for SoyChlor (SC, West Central Soy; Appendix C) and NRC (1998) values for corn (C) and soybean meal (SBM). Proximate analysis and NDF were conducted on SC for calculation of ME. Metabolizable energy was calculated using Noblet's equations (Appendix B). The equation that was used to determine DCAD levels in the diets is in Appendix B. The diets were formulated to 3,300 kcal/kg ME and 1.02% Lys. The diets met or exceeded 105% of the requirement (NRC, 1998) for lactating sows anticipating no lactation weight loss with pigs gaining 250 g/d. Dietary treatments (Table 3.1) for Exp. 1 consisted of a positive control (PC) C-SBM diet and a C-SBM diet plus 1.5, 2.5, and 3.5% SC to achieve DCAD of 140, 99, 75, and 45 mEq/kg. In Exp. 2, dietary treatments consisted of the same PC C-SBM (140 mEq/kg) and C-SBM diet with reduced DCAD (45 mEq/kg). Sows were weighed and moved into the farrowing house on their respective date of gestation, and from this point forward, the treatment diets were started and continued through weaning. Upon entering the farrowing house until d 1 postfarrowing, the sows were fed approximately 2.5 kg/d (as-fed basis). On d 1 postfarrowing, sows and feed containers were weighed to determine weight change per day and ADFI from d 107 of gestation in Exp.1 and d 111 of gestation in Exp. 2 to d 1 postfarrowing.

The farrowing house is an environmentally controlled building with 28 individual farrowing crates and an under-floor flush system. Each farrowing crate is 1.5 x 2.1 m, with a cast iron-floor for the sow and plastic slotted floor for the pigs. Each crate contains a stainless steel feeder and nipple waterer for the sows and a nipple waterer for the pigs. Within 24 h of farrowing, litters were weighed, ear-notched, given a 1-mL shot of iron dextran (Phoenix Scientific Inc., St. Joseph, MO), umbilical cords were sprayed with iodine, and needle teeth were clipped.

During processing, litters were also adjusted by cross-fostering within respective treatments to approximately 10 pigs per litter if necessary. From d 1 postfarrowing to weaning, the sows were fed three times daily. In the four farrowing groups, pigs were weaned at an average of 19, 18, 16, and 15 d. All pigs were weaned on the same day regardless of their farrowing date. At weaning, the sows and feed containers were weighed to determine lactation weight change per day and lactation ADFI from d 1 postfarrowing to weaning. Within 2 d of weaning, sows were checked daily for signs of estrus. In Exp. 1, 7 sows did not return to estrus (one from PC; six from the reduced DCAD diets) within 7 d postweaning. In Exp. 2, there were six sows fed the PC and two sows fed the reduced DCAD diet that did not return to estrus within 7 d postweaning.

Blood Collection

On d 1 postfarrowing, approximately 6 mL of blood was collected from the anterior vena cava with a 16 gauge needle and placed into a 10-mL tube (BD Vacutainer, Franklin Lakes, NJ) that contained no additive. The sample of blood was immediately centrifuged in an Allegra 6R Centrifuge (Beckman Coulter, Inc., Palo Alto, CA.) at 1,500 *g* at 4°C for 25 min. After centrifugation, the plasma was poured into 5 mL tubes and stored at 20°C until analysis. The plasma samples were analyzed using flame atomic absorption spectrophotometry (Perkin Elmer Analyst 300) after a 1:100 dilution with 0.5% lanthanum oxide. For Exp. 1, the samples were analyzed once whereas, in Exp. 2, samples were analyzed twice and then averaged to get a mean for plasma Ca concentration.

Urine Collection

A midstream urine sample was collected from each sow for pH determination when sows averaged 11 ± 2 d postfarrowing in Exp. 1 and 12 ± 3 d postfarrowing in Exp. 2. Two urine samples were collected from each sow and averaged together on two consecutive days. Each urine collection period lasted 3 d. The pH was determined by a pH/mV/thermometer with an attached islet sensor pH probe.

Sow Response Variables

The sow response variables included weight at d 107 and d 111 of gestation,

Table 3.1. Composition of experimental diets, as-fed basis

Item	Dietary cation-anion difference, mEq/kg ^a			
	140 ^{b,c}	99 ^b	75 ^b	45 ^{b,c}
Ingredient, %				
Corn	66.007	65.081	64.464	63.848
Soybean meal	28.090	27.788	27.586	27.384
SoyChlor 16-7 ^a	-----	1.500	2.500	3.500
Monocalcium phosphate	2.006	1.999	1.995	1.990
Limestone	1.528	1.400	1.315	1.229
Dry fat	1.168	1.032	0.940	0.849
Vitamin premix ^d	0.500	0.500	0.500	0.500
Salt	0.500	0.500	0.500	0.500
Choline chloride, 60%	0.100	0.100	0.100	0.100
Trace mineral premix ^e	0.100	0.100	0.100	0.100
Calculated composition:				
ME, kcal/kg	3,300	3,300	3,300	3,300
Lysine, %	1.020	1.020	1.020	1.020
Methionine, %	0.300	0.302	0.303	0.304
Tryptophan, %	0.222	0.221	0.220	0.220
Sodium, %	0.216	0.218	0.219	0.219
Potassium, %	0.823	0.822	0.822	0.821
Chloride, %	0.350	0.493	0.587	0.681
Sulfur, %	0.210	0.217	0.221	0.225
Calcium, %	1.000	1.000	1.000	1.000
Magnesium, %	0.204	0.199	0.197	0.194
Phosphorus, %	0.800	0.800	0.800	0.800
DCAD ^f	140	99	75	45

^aDietary cation-anion difference was achieved by adding SoyChlor at 0, 1.5, 2.5, and 3.5% of the diet, respectively.

^bDietary treatments fed in Exp. 1.

^cDietary treatments fed in Exp. 2.

^dProvided the following per kilogram of diet: vitamin A, 11,023 IU; vitamin D₃, 3,307 IU; vitamin E, 88 IU; menadione, 8.3 mg; riboflavin, 13 mg; Ca-d-pantothenic acid, 50 mg; niacin, 88 mg; vitamin B₁₂, 61 µg; d-biotin, 441 µg; folic acid, 3.3 mg; pyridoxine, 4.4 mg; thiamin, 4.4 mg; and vitamin C, 110 µg.

^eProvided the following per kilogram of diet: zinc, 127 mg; iron, 127 mg; manganese, 22 mg; copper, 12.7 mg; iodine, 0.80 mg; and selenium, 0.3 mg.

^fDCAD = ((Na + K + 0.15Ca + 0.15Mg) – (Cl + 0.20S + 0.30P)).

^gME, kcal/kg = DE x (1.003 – (0.0021 x % CP)).

d 1 postfarrowing, and at weaning; weight change per day from prefarrowing, lactation, and overall; ADFI, ADG, and G:F prefarrowing, during lactation, and overall; lactation length, days to estrus, total number pigs born, pigs born alive, stillbirths, and mummies.

Litter Response Variables

The litter response variables were live and total birth weights, number weaned, number nursed, initial litter weight after cross-fostering, initial litter weight adjusted for mortality, final litter weight, litter weight gain, and percent survivability.

Statistical Analysis

Data were analyzed by ANOVA procedures appropriate for a randomized complete block design (Steel and Torrie, 1980) using the GLM procedures of SAS (SAS Inst. Inc., Cary, NC). All data were analyzed with the sow as the experimental unit. In both experiments, covariates were used to analyze some response variables, but orthogonal contrast statements (linear and quadratic) were also used (Exp. 1 only) to analyze treatment differences. There were no covariates used for the following data: sow 107 and 111 d weight, G:F from d 107 and 111 of gestation to d 1 postfarrowing, during lactation, and overall; lactation length, total number pigs born, pigs born alive, stillbirths, mummies, live and total birth weights, initial litter weight after cross-fostering, and initial litter weight adjusted for mortality. Sow d 107 and 111 of gestation weight was used as a covariate for sow d 1 postfarrowing and weaning weight; prefarrowing, lactation, and overall weight change per day; ADG from d 107 and 111 of gestation to d 1 postfarrowing, during lactation, and overall, and ADFI from d 107 and 111 of gestation to d 1 postfarrowing. Sow d 111 weight affected ($P < 0.02$) ADFI from d 111 of gestation to d 1 postfarrowing. Lactation length was used as a covariate for ADFI during lactation and overall, number of pigs nursed, final litter weight, litter weight gain, days to estrus, and percent survivability. Initial litter weight after cross-fostering and lactation length were covariates for final litter weight and litter weight gain. Neither of the covariates were significant ($P > 0.10$) for the response variables final litter weight and litter weight gain. Number of pigs nursed was used as a covariate for number of pigs weaned. For both experiments, an alpha level of 0.10 was used to

indicate significant treatment differences.

Results

Experiment 1

Experiment 1 was a preliminary study conducted to determine the optimum level of SC needed to change DCAD to reduce urinary pH. The results from Exp. 1 indicate that DCAD had little effect on sow (Table 3.2) and litter response (Table 3.3) variables. As DCAD decreased, ADFI during lactation ($P < 0.08$) and overall ($P < 0.06$) increased linearly. As DCAD decreased in the diets, urinary pH linearly decreased ($P < 0.001$). Reducing DCAD tended to linearly increase ($P = 0.15$) plasma Ca concentrations. The results indicate that changing DCAD to 45 mEq/kg had the greatest decrease in urine pH (Table 3.4), and this DCAD was used in Exp. 2.

Experiment 2

Sow Response Variables. In Exp. 2 (Table 3.5), ADFI from d 111 of gestation to d 1 postfarrowing decreased as DCAD decreased ($P < 0.02$). However, ADFI during lactation and overall were not affected ($P > 0.10$) by DCAD. Neither ADG nor G:F were affected by DCAD ($P > 0.10$). Sows fed the reduced DCAD diet tended to have reduced weight change during prefarrowing (-18.71 vs. -15.11), lactation (-8.67 vs. -6.84), and overall (-27.42 vs. -21.94), but the effects were not significant ($P > 0.10$). Sows fed the PC diet had decreased days to estrus compared to sows fed the reduced DCAD diet ($P < 0.04$). The DCAD diet did not affect total number pigs born, pigs born alive, stillbirths, or mummies ($P > 0.10$).

Litter Response Variables. Reducing DCAD (Table 3.6) tended to result in a higher number of pigs weaned (8.79 vs. 8.43), but the effect was not significant ($P = 0.18$). Reducing DCAD tended to increase initial litter weight adjusted for mortality, but the effect was not significant ($P = 0.13$). Also, percent survivability of pigs nursing from sows fed the reduced DCAD diet tended to be higher (89.42 vs. 85.77), but the effect was not significant ($P = 0.19$). Feeding reduced DCAD diets did not affect live and total birth weights, number nursed, initial litter weight after cross-fostering, final litter weight, or litter weight gain ($P > 0.10$). Sows fed reduced DCAD ($P < 0.001$) had decreased urinary

Table 3.2. The effects of dietary cation-anion difference on sow response variables, Experiment 1^a

Response variables	Dietary cation-anion difference, mEq/kg ^b				SEM	P = F		
	140	99	75	45		Linear	Quadratic	P value
Parity	1.50	2.00	2.33	2.00	0.58	0.46	0.48	
d 107 wt, kg	252.65	266.56	268.75	267.17	18.78	0.58	0.70	
24h wt, kg ^{i,k}	247.00	252.54	258.05	251.48	5.94	0.44	0.47	0.57
Weaning wt, kg ^{i,k}	241.22	246.76	252.54	244.36	6.78	0.48	0.46	0.64
Pre-farr BW change, kg ^{f,i,k}	-16.20	-10.66	-5.15	-11.71	5.94	0.75	0.54	0.57
Lactation BW change, kg ^{g,i,k}	-5.78	-5.78	-5.51	-7.12	3.12	0.81	0.78	0.98
Overall BW change, kg ^{h,i,k}	-21.98	-16.44	-10.65	-18.83	6.78	0.85	0.50	0.78
ADG 1, kg ^{c,i,k}	-1.76	-1.23	-0.54	-1.75	0.59	0.92	0.36	0.38
ADG 2, kg ^{d,i,k}	-0.33	-0.32	-0.33	-0.35	0.16	0.94	0.89	0.99
OADG, kg ^{e,i,k}	-0.81	-0.61	-0.36	-0.69	0.24	0.79	0.44	0.56
ADFI 1, kg ^{c,i,k}	2.34	2.34	2.63	2.40	0.12	0.51	0.46	0.32
ADFI 2, kg ^{d,j,k}	5.03	5.40	6.31	6.09	0.53	0.08	0.37	0.19
OADFI, kg ^{e,j,k}	4.16	4.44	4.85	5.00	0.39	0.06	0.84	0.29
G:F 1, kg ^c	-0.660	-0.566	-0.302	-0.801	0.29	0.90	0.35	
G:F 2, kg ^d	-0.060	-0.059	-0.049	-0.059	0.03	0.93	0.86	
OG:F, kg ^e	-0.178	-0.140	-0.083	-0.152	0.06	0.60	0.38	
Lactation length, d	19.33	18.33	18.00	21.20	0.90	0.20	0.04	
Days to estrus, d ^{j,k}	4.21	4.71	4.70	4.49	0.48	0.93	0.22	0.77
Total pigs born	11.33	11.00	11.50	12.00	1.73	0.74	0.82	
Pigs born alive	10.67	9.00	9.00	10.20	1.36	0.82	0.33	
Stillbirths	0.67	2.00	1.83	0.80	0.64	0.93	0.10	
Mummies	0.00	0.00	0.67	1.00	0.26	0.005	0.54	

^an = 6, 3, 6, and 5 for 140, 99, 75, and 45 dietary cation-anion difference, respectively.^bDietary cation-anion difference was achieved by adding SoyChlor at 0, 1.5, 2.5, and 3.5% of the diet, respectively.^cADG 1, ADFI 1, and GF 1 are measurements taken from d 107 until d 1 postfarrowing.^dADG 2, ADFI 2, and GF 2 are measurements taken from d 1 postfarrowing until weaning.^eOADG, OADFI, and OGF are measurements taken from d 107 until weaning.^fPre-farrowing BW change = the difference in sow wt at d 1 postfarrowing and sow wt at d 107 of gestation.^gLactation BW change = the difference in sow weaning wt and sow wt at d 1 postfarrowing.^hOverall BW change = the difference in sow weaning wt and sow wt at d 107 of gestation.ⁱTreatment means were reported using sow wt at d 107 of gestation as a covariate.^jTreatment means were reported using lactation length as a covariate.^kP values are only reported for the response variables that used covariates.

Table 3.3. The effects of dietary cation-anion difference on litter response variables, Experiment 1^a

Response variables	Dietary cation-anion difference, mEq/kg ^b					P = F		
	140	99	75	45	SEM	Linear	Quadratic	P value
Birth wt live, kg	16.51	14.64	13.33	16.49	1.90	0.87	0.22	
Birth wt total, kg	17.26	17.13	15.78	17.67	2.16	0.99	0.66	
Nursed ^{f,h}	10.46	9.39	9.20	9.36	1.26	0.81	0.22	0.82
Weaned ^j	8.75	8.24	9.03	8.52	0.46	0.84	0.26	0.70
Initial litter wt, kg (ACF) ^c	16.80	14.64	13.15	16.41	1.69	0.72	0.14	
Initial litter wt, kg (AM) ^d	14.81	12.81	12.37	14.67	1.61	0.90	0.22	
Final litter wt, kg ^j	44.01	45.27	49.16	49.96	4.65	0.37	0.17	0.71
Litter wt gain, kg/d ^{e,i}	1.58	1.61	1.81	1.85	0.21	0.40	0.38	0.69
Survival, % ^h	90.22	84.69	94.71	89.32	4.97	0.77	0.89	0.55

^an = 6, 3, 6, and 5 for 140, 99, 75, and 45 dietary cation-anion difference, respectively.

^bDietary cation-anion difference was achieved by adding SoyChlor at 0, 1.5, 2.5, and 3.5% of the diet, respectively.

^cACF = after cross-fostering.

^dAM = initial litter weights were adjusted for mortality once cross-fostering had occurred.

^eLitter wt gain = (final litter wt – initial litter wt (AM)) / lactation length.

^fNursed = the number of pigs the sow nursed after cross-fostering occurred.

^gSurvival % = pigs weaned / pigs nursed * 100.

^hTreatment means were reported using lactation length as a covariate.

ⁱTreatment means were reported using initial litter wt (ACF) as a covariate.

^jTreatment means were reported using nursed as a covariate.

Table 3.4. The effects of dietary cation-anion difference on urinary pH and plasma Ca concentrations, Experiment 1^a

Response variables	Dietary cation-anion difference, mEq/kg ^b					P = F	
	140	99	75	45	SEM	Linear	Quadratic
Urinary pH ^c	7.28	7.13	6.85	6.21	0.17	0.001	0.23
Plasma Ca, mg/dL	7.70	7.70	7.97	7.98	0.16	0.15	0.97

^an = 6, 3, 6, and 5 for 140, 99, 75, and 45 dietary cation-anion difference, respectively.

^bDietary cation-anion difference was achieved by adding SoyChlor at 0, 1.5, 2.5, and 3.5% of the diet, respectively.

^cUrinary pH = an average of two urine samples were taken on two consecutive days from each sow 11 ± 2 d postfarrowing.

Table 3.5. The effects of dietary cation-anion difference on sow response variables, Experiment 2^a

Response variables	Covariates	Dietary cation-anion difference, mEq/kg ^b			
		140	45	SEM	<i>P</i> value
Parity		1.45	1.58	0.26	0.74
d 111 wt, kg		261.85	270.18	6.25	0.35
24h wt, kg	Sow d 111 wt	247.42	251.01	1.98	0.21
Weaning wt, kg	Sow d 111 wt	238.62	244.08	2.51	0.13
Pre-farr BW change, kg ^f	Sow d 111 wt	-18.71	-15.11	1.98	0.21
Lactation BW change, kg ^g	Sow d 111 wt	-8.67	-6.84	1.94	0.51
Overall BW change, kg ^h	Sow d 111 wt	-27.42	-21.94	2.51	0.13
ADG 1, kg ^c	Sow d 111 wt	-3.50	-3.07	0.48	0.53
ADG 2, kg ^d	Sow d 111 wt	-0.51	-0.44	0.12	0.67
OADG, kg ^e	Sow d 111 wt	-1.25	-1.00	0.12	0.13
ADFI 1, kg ^c	Sow d 111 wt	2.80	2.06	0.22	0.02
ADFI 2, kg ^d	Lactation length	5.04	5.55	0.24	0.15
OADFI, kg ^e	Lactation length	4.44	4.65	0.19	0.43
G:F 1, kg ^c		-1.88	-2.20	0.53	0.67
G:F 2, kg ^d		-0.11	-0.09	0.03	0.58
OG:F, kg ^e		-0.31	-0.24	0.04	0.18
Lactation length, d		16.24	16.30	0.46	0.93
Days to estrus, d	Lactation length	4.19	4.58	0.12	0.04
Total number pigs born		12.09	11.64	0.62	0.61
Pigs born alive		10.15	10.18	0.61	0.97
Stillbirths		1.55	1.27	0.22	0.39
Mummies		0.39	0.18	0.13	0.25

^aData are means of 33 sows per treatment over three farrowing groups.^bDietary cation-anion difference was achieved by adding SoyChlor at 0 and 3.5% of the diet.^cADG 1, ADFI 1, and GF 1 are measurements taken from d 111 until farrowing.^dADG 2, ADFI 2, and GF 2 are measurements taken on d 1 postfarrowing until weaning.^eOADG, OADFI, and OGF are measurements taken from d 111 until weaning.^fPre-farrowing BW change = the difference in sow wt at d 1 postfarrowing and sow wt at d 111 of gestation.^gLactation BW change = the difference in sow weaning wt and sow wt at d 1 postfarrowing.^hOverall BW change = the difference in sow weaning wt and sow wt at d 111 of gestation.

Table 3.6. The effects of dietary cation-anion difference on litter response variables, Experiment 2^a

Response variables	Covariates	Dietary cation-anion difference, mEq/kg ^b			
		140	45	SEM	<i>P</i> value
Birth wt live, kg		15.15	15.36	0.81	0.85
Birth wt total, kg		17.30	16.94	0.78	0.75
Weaned	Nursed	8.43	8.79	0.19	0.18
Nursed ^f	Lactation length	9.61	10.02	0.30	0.33
Initial litter wt, kg (ACF) ^c		14.63	15.44	0.54	0.29
Initial litter wt, kg (AM) ^d		12.77	13.98	0.56	0.13
Final litter wt, kg	Lact d / ILW(ACF)	44.55	45.24	1.24	0.70
Litter wt gain, kg ^e	Lact d / ILW(ACF)	1.91	1.96	0.07	0.62
Survival % ^g	Lactation length	85.77	89.42	1.95	0.19

^aData are means of 33 sows per treatment over three farrowing groups.

^bDietary cation-anion difference was achieved by adding SoyChlor at 0 and 3.5% of the diet.

^cACF = after cross-fostering.

^dAM = initial litter weights adjusted for mortality once cross-fostering had occurred.

^eLitter wt gain = (final litter wt – initial litter wt (AM)) / lactation length.

^fNursed = the number of pigs the sow nursed after cross-fostering occurred.

^gSurvival % = pigs weaned / pigs nursed * 100.

Table 3.7. The effects of dietary cation-anion difference on sow response variables in subsequent farrowings, Experiment 2

Dietary cation-anion difference, mEq/kg ^a				
Response variables	140	45	SEM	<i>P</i> value
n	27	21		
Total number pigs born	9.63	11.48	0.73	0.08
Pigs born alive	8.26	10.38	0.59	0.02
Stillbirths	1.00	0.86	0.26	0.71
Mummies	0.41	0.26	0.16	0.59
Mortality ^b	0.61	0.97	0.18	0.19
Birth wt live, kg ^c	14.32	16.09	1.21	0.31
Birth wt total, kg ^d	15.69	17.07	1.21	0.43

^aDietary cation-anion difference was achieved by adding SoyChlor at 0 and 3.5% of the diet and was fed during the previous lactation.

^bMortality was analyzed using pigs born alive as a covariate.

^cBirth wt live = is the wt of all pigs born alive.

^dBirth wt total = is the wt of all pigs born alive and dead.

Table 3.8. The effects of dietary cation-anion difference on urinary pH and plasma Ca concentrations, Experiment 2^a

Response variables	Dietary cation-anion difference, mEq/kg ^b			<i>P</i> value
	140	45	SEM	
Urinary pH ^c	7.03	5.66	0.07	<.0001
Plasma Ca, mg/dL ^d	9.00	8.92	0.20	0.65

^aData are means of 33 sows per treatment over three farrowing groups.

^bDietary cation-anion difference was achieved by adding SoyChlor at 0 and 3.5% of the diet.

^cUrinary pH = an average of two urine samples taken on two consecutive days from each sow 12 ± 3 d postfarrowing.

^dPlasma Ca concentration is the mean from a sample that was analyzed two times.

pH compared to sows fed the PC diet (Table 3.8). Plasma Ca concentrations were not affected ($P > 0.10$) by DCAD (Table 3.8).

There were 27 sows fed the PC diet and 21 sows fed the reduced DCAD diet that were evaluated during their next farrowing (Table 3.7). The sows that were fed the reduced DCAD diet during the previous lactation had an increased total number of pigs born ($P < 0.08$) and pigs born alive ($P < 0.02$). Reducing DCAD did not affect stillbirths, mummies, mortality, or live and total birth weights ($P > 0.10$).

Discussion

Experiment 1

The results from Exp. 1 showed that feeding diets with reduced DCAD had little effect on sow and litter response variables. However, ADFI during lactation and overall was linearly increased as DCAD decreased in the diet. Yen et al. (1981) fed finishing pigs 4% CaCl_2 , which decreased ADFI. Our results indicate a linear ($P < 0.001$) decrease in urine pH as DCAD is decreased in the diet. Urinary pH is the response that we used to determine the percentage of SC that would be fed in Exp. 2.

Experiment 2

Changing DCAD reduced ADFI from d 111 of gestation to d 1 postfarrowing. However, DCAD had no effect on ADFI during lactation and overall. DeRouchey et al. (2003) conducted a pilot study where he fed DCAD of -200, -100, 0, 100, and 200 mEq/kg. The results showed that negative DCAD reduced feed intake, and the lowest DCAD that should be fed to lactating sows was 0 mEq/kg. After the pilot study, DCAD's of 0, 100, 200, 350, and 500 mEq/kg were fed. The authors reported that increasing DCAD had no effect on ADFI (DeRouchey et al., 2003). Research in dairy cows and in growing pigs suggests that feeding diets with a negative DCAD decreases feed intake (Escobosa et al., 1984; Patience et al., 1987). DeRouchey et al. (2003) reported that increasing DCAD reduced percent survivability of pigs and number weaned. In our experiment, results indicate that reducing DCAD (45 mEq/kg) tended to improve percent survivability of pigs and number weaned, but the effect was not significant.

DeRouchey et al. (2003) reported that when DCAD is reduced below a standard C-SBM diet, sow's milk production increases, which may increase percent survivability in pigs. Overall, our results and those reported by DeRouchey et al. (2003) are in agreement. Dove and Haydon et al. (1994) fed sows diets with DCAD levels of 130.8, 161.2, and 250.8 mEq/kg and reported no differences in feed intake, days to estrus, percent survivability in pigs or litter weight gain regardless of dietary treatments. In our experiment, reducing DCAD significantly increased the number of days to estrus. Our results cannot be compared with those of Dove and Haydon et al. (1994) because they did not decrease acid-base balance below 130 mEq/kg. In our experiment, the highest DCAD was 140 mEq/kg.

Acidifiers are not growth promotants. They are included in the diet for metabolic purposes such as bone metabolism, Ca metabolism, maintaining blood pH, and reducing urinary pH. DeRouchey et al. (2003) reported that increasing DCAD linearly increased urinary pH and bacterial counts in the urine. Sows with reduced bacterial concentrations had a reduced risk of urinary tract infections. Sows that farrow in areas contaminated with excess fecal matter are at risk for bacterial infections. Bacterial infections may decrease rebreeding performance and litter health. DeRouchey et al. (1998) reported that Ca concentrations in the blood decreased as DCAD increased. This response does not agree with our results. Our data showed that plasma Ca slightly decreased as DCAD decreased, but the effect was not significant. Jackson et al. (1994) reported that plasma Ca is similar among treatments regardless of DCAD or Ca% in the diet. Tucker et al. (1988) and Waterman et al. (1991) used dairy cows to investigate plasma Ca in response to high and low DCAD diets and found no relationship between the two. These observations agree with our results.

CHAPTER 4

SUMMARY AND CONCLUSIONS

This research was conducted to determine if changing DCAD of a standard C-SBM diet during late gestation and in lactation would have an effect on sow productivity.

Experiment 1 was conducted to determine what percentage of acidifier would be adequate to reduce urinary pH. Reducing DCAD increased the number of stillbirths ($P < 0.10$). Reducing the DCAD linearly decreased urinary pH ($P < 0.001$).

In Exp. 2, changing DCAD did not affect ($P > 0.10$) sow growth or litter response variables. However, DCAD decreased ($P < 0.001$) urinary pH. Reducing DCAD (45 mEq/kg) below a standard C-SBM diet (140 mEq/kg) tended to improve sow and litter response variables. Reducing DCAD tended to improve number of pigs weaned ($P = 0.18$), initial litter weight adjusted for mortality ($P = 0.13$), and percent survivability ($P = 0.19$), but these effects were not significant. In our experiment, reducing DCAD during the previous lactation improved some response variables in subsequent farrowings in sows. Sows that had been fed a reduced DCAD (45 mEq/kg) diet showed an increase in total number of pigs born ($P < 0.08$) and pigs born alive ($P < 0.02$).

In conclusion, decreasing cations and increasing anions can improve sow health and reproductive performance. These experiments showed that reducing DCAD reduced urine pH and some sow and litter performance variables were improved when compared to the other dietary treatments. Long term studies need to be conducted to determine if reducing DCAD throughout gestation would increase sow productivity.

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APPENDIX A

List of Abbreviations

Item	Abbreviation
Adjusted for mortality	AM
After cross-fostering	ACF
Corn	C
Dietary Cation-Anion Difference	DCAD
Metabolizable Energy	ME
Soybean meal	SBM
SoyChlor	SC

APPENDIX B

Equations Used in Diet Formulations

Equations for use in estimating energy from chemical analysis of a feedstuff (NRC, 1998)

$$GE = 4,143 + (56 \times \% EE) + (15 \times \% CP) - (44 \times \% Ash)$$

$$DE = 949 + (0.789 \times GE) - (43 \times \% Ash) - (41 \times \% NDF)$$

With the following correction for increased body weight:

$$DE = 1,391 + (0.58 \times DE) + (23 \times \% EE) + (12.7 \times \% CP)$$

$$ME = DE \times (1.003 - (0.0021 \times \% CP))$$

DCAD Equation (Philip Jardon, DVM, MS):

$$(Na + K + .15 Ca + .15 Mg) - (Cl + .20 S + .30 P) = DCAD \text{ in mEq/kg DM}$$

APPENDIX C

Analysis of SoyChlor (SC)

Item	%
Crude protein	22.10
Crude fat	6.81
Crude fiber	4.90
Ash	16.14
Moisture	10.77
NDF	18.47
Taurine	0.04
Hydroxyproline	0.08
Aspartic acid	1.52
Threonine	0.84
Serine	0.99
Glutamic acid	3.50
Proline	1.65
Lanthionine	0.01
Glycine	0.83
Alanine	1.51
Cysteine	0.39
Valine	1.12
Methionine	0.34
Isoleucine	0.85
Leucine	2.43
Tyrosine	0.80
Phenylalanine	1.07
Hydroxylysine	0.00
Histidine	0.58
Ornithine	0.02
Lysine	0.77
Arginine	0.98
Tryptophan	0.10
Ca	3.40
Mg	2.73
P	0.41
K	0.56
Na	0.11
S	0.61
Cl	9.49

VITA

Melanie L. Roux was born April 22, 1980, in New Orleans, Louisiana. Melanie grew up in New Orleans until the age of 15, and then she moved to Ponchatoula, Louisiana. She graduated from Ponchatoula High School in May 1998. Melanie began her undergraduate career at Louisiana State University where she earned a Bachelor's Degree in Animal Science in December 2002. In August of 2003, Melanie returned to Louisiana State University in pursuit of a master of science degree in non ruminant nutrition. Currently, Melanie is a candidate for her master's degree in animal sciences- non-ruminant nutrition.